# **COLLEGE OF BIOTECHNOLOGY**

# SYLLABUS-III

# B. TECH. (BIOTECHNOLOGY)

(As per V- Dean Committee Recommendations)



# SARDAR VALLABH BHAI PATEL UNIVERSITY OF AGRICULTURE AND TECHNOLOGY, MEERUT- 250 110

Syallabus I: 2003-04 to 2011-12.

Syallabus II: Revision- 2012-13 to 2015-16.

Syallabus III: Revision (as per V Dean Committee) 2016-17 onwards.

Reprint: March-2018

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Non-Teaching: Mr. Harish Chandrra Mrs. Pooja Aggarwal Mrs. Suchi Gupta

Printed at :-

Semester	: Course No.	Name of Course	Credits	Page No.
Sem. I	BAS 112	Human Ethics	1 (1-0-0)	13
	BAS 116	Basic Mathematics-I	3 (3-0-0)	13
	BAS 114	Basic Botany	3 (2-0-1)	14
	BAS 115	Communication Skills & Personality Development	2 (1-0-1)	15
	AGE/ SAC/	Environmental Studies and Disaster Management	3 (2-0-1)	15
	AGR 114			
	AGE 113	Food Science and Processing	2 (1-0-1)	17
	AGR 113	Crop Production Technology	3 (2-0-1)	18
	BTM 111	Basic Genetics	3 (2-0-1)	19
	BTO 111	Introduction to Biotechnology	3 (2-0-1)	20
	BTC 111	Cell Biology	2 (2-0-0)	20
	NCC/NSS/	NCC/NSS/PEY (NG)	1 (0-1-0)	
	PEY-111			
		(R: BAS 116 or BAS 114) Total	23 (18-0-8)	
Sem. II	BAS 121	Basic Zoology	3 (2-0-1)	21
	BAS 122	Basic Mathematics- II	3 (3-0-0)	21
	BAS 123	Basic Statistics	2 (1-0-1)	22
	HOR 121	Production Technologies for Horticultural Crops	3 (2-0-1)	23
	VAN/VPB 123	Anatomy and Physiology of Livestock	3 (3-0-0)	24
	GPB 122	Basics of Plant Breeding	3 (2-0-1)	24
	AAP 121	Introduction to Animal Breeding	3 (2-0-1)	25
	BTO 121	Plant Tissue Culture	3 (2-0-1)	26
	BTM 121	Molecular Biology	3 (2-0-1)	26
	BTC 121	Biodiversity and its conservation	2 (2-0-0)	27
	BTP 121	Microbiology	3 (2-0-1)	28
	(R: BAS 1	21 or BAS 122, OP: HOR 121 or VPB 123, GPB 12	2 or AAP 121)	)
		Total	22 (23-0-9)	

## Semester wise Distribution of Courses

			Total	22 (23-0-9)	
Sem. III	BAS 211	Biomathematics		3 (2-0-1)	29
	BAS 214	Information and Communication Tec	hnology	2 (1-0-1)	29

	AAE 212	Economics and Marketing	3 (2-0-1)	30
	ENT/ PPA 212	Fundamentals of Crop Protection	3 (2-0-1)	31
	LPT 211	Livestock Product Technology	3 (2-0-1)	32
	GPB 212	Breeding of Field Crops	3 (2-0-1)	33
	VMD/VMC 212	Animal Health Care	3 (2-0-1)	34
	LPM 212	Livestock Production and Management	3 (2-0-1)	34
	BTR 211	Recombinant DNA Technology	3 (2-0-1)	35
	BTB 211	Plant Physiology	3 (2-0-1)	36
	NCC/NSS/	NCC/NSS/PEY (NG)	1 (0-1-0)	
	PEY-111			
	(OP: ENT 212	2 or LPT 211, GPB 212 or VMD 211) Total	24 (15-0-9)	
Sem. IV	BAS 221	Biophysics	3 (2-0-1)	36
	AEC/ AAE 222	Entrepreneurship Development and Business		
		Management	2 (1-0-1)	37
	BTB/ AGE 221	Electronics and Instrumentation in Biotechnology	2 (1-0-1)	38
	BTB 222	General Biochemistry	4 (3-0-1)	39
	BTI/ BTR 221	Introductory Bioinformatics	3 (2-0-1)	40
	BTM 221	Plant Genetic Transformation	3 (2-0-1)	41
	BTC 221	Classical and Molecular Cytogenetics	3 (2-0-1)	41
	BTP 221	Microbial Genetics	3 (2-0-1)	42
		Total	23 (15-0-8)	
Sem. V	BAS 311	Agricultural Informatics	3 (2-0-1)	43
	BTI/ VPB/			
	VMC 312	Animal Biotechnology	4 (3-0-1)	44
	BTI 311	Immunology	3 (2-0-1)	45
	BTM 311	Molecular Genetics	2 (2-0-0)	45
	BTM 312	Nanobiotechnology	2 (2-0-0)	46
	BTF 311	Molecular Marker Technology	2 (2-0-0)	46
	BTF 312	IPR, Biosafety and Bioethics	2 (2-0-0)	47
	BTR 311	Genomics and Proteomics	3 (3-0-0)	47
	BTB 311	Enzymology & Enzyme Technologies	3 (2-0-1)	48
		Total	24 (20-0-4)	

Sem. VI	BAS 321	Biostatistics	3 (2-0-1)	49
	BTI/BTR 321	Computational Biology	3 (2-0-1)	49
	Optional/ Electiv	re Course (04 no.)		
	1.	Plant Biotechnology	18 (12-0-6)	50
	2.	Animal Biotechnology	18 (13-0-5)	55
	3.	Microbial and Environmental Biotechnology	18 (14-0-4)	59
	4.	Bioinformatics	18 (11-0-7)	63
		Total	24 (15/18 -0-	· 6/9)
Sem. VII	BTS 411	Educational Tour (NG)	1 (0-1-0)	67
	BTS 412	Student READY – (04 Optional topics)		
		In-house Skill Development Modules	20 (0-0-20)	67
	BTO/AGE 411	Bioprocess Engineering, Bioseperation and		
		Downstream processing	3 (3-1-0)	68
		Total	24	
Sem. VII	<b>I</b> BTS 421	Student READY - Project Formulation,		
		Execution and Presentation	10	69
	BTS 422	Student READY - Entrepreneurial Development		
		in Biotechnology	10	69
		Total	20	
		Grand Total	184 (59 Cou	rses)

<b>S.</b>	Semester							Total Credits	
	Ι	II	III	IV	V	VI	VII	VIII	
Credit	23	22	24	23	24	24	24	20	184 (59 Courses)

Elective I. Plant Biotechnology					
Course No.	Course Title	Credit hours			
BTF 321	Plant Tissue Culture and its Applications	3 (2-0-1)			
BTM 321	Principles and Applications of Plant Genetic Transformation	3 (2-0-1)			
BTM 322	Epigenetics and Gene Regulation	3 (2-0-1)			
BTR 322	Applications of Genomics and Proteomics	3 (2-0-1)			
GPB 322	Molecular Breeding in Field Crops	3 (2-0-1)			
HOR 322	Molecular Breeding of Horticultural Crops and Forest Trees	3 (2-0-1)			
Elective II. Animal Bio	technology	1			
VPB 321	Animal Genomics	3 (2-0-1)			
VPB/VGO 322	Embryo Transfer Technologies	3 (2-0-1)			
BTI/VPB/VMC 323	Principles and Procedures of Animal Cell Culture	3 (2-0-1)			
BTI/VPB 324	Transgenic Animal Production	3 (3-0-0)			
BTI/VPB 325	Molecular Diagnostics	3 (2-0-1)			
BTI/VMC 326	Molecular Virology and Vaccine Production	3 (2-0-1)			
Elective III. Microbial	and Environmental Biotechnology	1			
BTP 321	Microbial Biotechnology	3 (2-0-1)			
BTP/BTR 324	Green Biotechnology	3 (2-0-1)			
BTC 321	Molecular Ecology and Evolution	3 (3-0-0)			
BTI/VPT 327	Molecular Pharming and Biopharmaceuticals	3 (2-0-1)			
BTO/AGE 321	Food Biotechnology	3 (2-0-1)			
BTO 322	Bio-prospecting of Molecules and Genes	3 (3-0-0)			
Elective IV. Bioinform	atics	•			
BAS 322	Programming for Bioinformatics	4 (2-0-2)			
BAS 323	Computational Methods for Data Analysis	2 (1-0-1)			
BTI/BTR 328	Bioinformatics Tools and Biological Databases	3 (2-0-1)			
BTI/BTR 329	Structural Bioinformatics	3 (2-0-1)			
BTI/VPT 330	Pharmacogenomics	3 (2-0-1)			
BTB/BAS 327	Metabolomics and System Biology	3 (2-0-1)			

Semester: VI Elective Courses in Biotechnology (any one to choose); Each Elective: Total Credit Hours=18 (Tentative)

E.

## (04 Optional topics) Total credits 20 (0-0-20)

(Tentative)

Elec	tive I. Plant Biotechnology	
S.	Course Title	Deptt.
1.	Plant Tissue Culture and its Applications	BTF
2.	Principles and Applications of Plant Genetic Transformation	BTM
3.	Epigenetics and Gene Regulation	BTM
4.	Applications of Genomics and Proteomics	BTR
5.	Molecular Breeding in Field Crops	GPB
6.	Molecular Breeding of Horticultural Crops and Forest Trees	HOR
Elec	tive II. Animal Biotechnology	
1.	Animal Genomics	VPB
2.	Embryo Transfer Technologies	VPB/VGO
3.	Principles and Procedures of Animal Cell Culture	BTI/VPB/VMC
4.	Transgenic Animal Production	BTI/VPB
5.	Molecular Diagnostics	BTI/VPB
6.	Molecular Virology and Vaccine Production	BTI/VMC
Elec	tive III. Microbial and Environmental Biotechnology	
1.	Microbial Biotechnology	BTP
2.	Green Biotechnology	BTP/BTR
3.	Molecular Ecology and Evolution	BTC
4.	Molecular Pharming and Biopharmaceuticals	BTI/VPT
5.	Food Biotechnology	BTO/AGE
6.	Bio-prospecting of Molecules and Genes	ВТО
Elec	tive IV. Bioinformatics	
1.	Programming for Bioinformatics	BAS
2.	Computational Methods for Data Analysis	BAS
3.	Bioinformatics Tools and Biological Databases	BTI/BTR
4.	Structural Bioinformatics	BTI/BTR
5.	Pharmacogenomics	BTI/VPT
6.	Metabolomics and System Biology	BTB/BAS

#### **Remedial Courses for I- Semester**

#### 1. Inter/ 10+2 Agriculture Group

BAS 114	Basic Botany	3 (2-0-1)	14
BAS 121	Basic Zoology	3 (2-0-1)	21
2. Inter/	/ 10+2 Biology Group		
BAS 116 BAS 122 <b>3. Inter</b> /	Basic Mathematics-I Basic Mathematics-II / 10+2 Maths. Group	3 (3-0-0) 3 (3-0-0)	13 21
BAS 114	Basic Botany	3 (2-0-1)	14
BAS 121	Basic Zoology	3 (2-0-1)	21

#### 4. Inter/ 10+2 (Bio+ Maths) Group None

- NG: Non Gradial course. R: Remedial Course.
- **OP:** Optional course; Students will select courses either from Plant or from Animal Sciences.
- NCC/NSS/PEY: National Cadet Corps/ National Service Scheme/ Physical Education and Yoga. These courses (two credits) will be of two years duration. One credit each will be offered during first semester and third semester respectively. Final result of these courses will be submitted after completion of two years.

#### **Examination and Evaluation.**

- External theory (50%)
- Internal Theory + Practical (50%)
- Courses with theory and practical

Mid-term Exam (30%) + Assignment (5%) in practical oriented courses + Practical (15%)

• Courses with only theory

Mid-term Exam (40%) + Assignment (10%)

• Courses with only practical:

(100%) Internal.

#### • Skill development Module and Project.

As given in respective sections.

S.	Department	Courses
	College of Agriculture	
1.	Basic Science	BAS 112, BAS 116, BAS 114, BAS 115, BAS121, BAS 122, BAS 123, BAS 211, BAS 214, BAS 221, BAS 311, BAS 321, BAS 322, BAS 323, BTB/BAS 327
2.	Agriculture Engineering & Food Technology	AGE 113, AGE/SAC/AGR 114, BTB/AGE 221, BTO/AGE 321, BTO/AGE 411
3.	Agronomy	AGE/SAC/AGR 114, AGR 113
4.	Soil Science	AGE/SAC/AGR 114
5.	Genetics & Plant Breeding	GPB 122, GPB 212, GPB 322
6.	Entomology	ENT/PPA 212
7.	Plant Pathology	ENT/PPA 212
8.	Horticulture	HOR 121, HOR 322
9.	Agriculture Economics	AAE 212, AEC/AAE 222
10.	Agriculture Extension	AEC/AAE 222
11.	Animal Production	AAP 121
	College of Veterinary and Au	nimal Science
1.	Veterinary Physiology and Biochemistry	VAN/VPB 123, BTI/VPB/VMC 312, VPB 321, VPB/VGO 322, BTI/VPB/VMC 323, BTI/VPB 324, BTI/VPB 325
2.	Livestock Production and Technology	LPT 211
3.	Veterinary Microbiology	VMD/VMC 212, BTI/VPB/VMC 312, BTI/VPB/VMC 323, BTI/VMC 326
4.	Livestock Production and Management	LPM 212
5.	Veterinary Medicine	VMD/VMC 212
6.	Veterinary Anatomy	VAN/VPB 123
7.	Veterinary Gynecology & Obstetrics	VPB/VGO 322
8.	Veterinary Pharmacology & Toxicology	BTI/VPT 327, BTI/VPT 330
	College of Biotechnology	
1.	Cell Biology	BTC 111, BTC 121, BTC 221, BTC 321.
2.	Physiology & Biochemistry	BTB 211, BTB/AGE 221, BTB 222, BTB 311,

# Department wise distribution of courses

		BTB/BAS 327.
3.	Molecular Biology &	BTM 111, BTM 121, BTM 221, BTM 311, BTE 312,
	Genetic Engineering	BTM 321, BTM 322.
4.	Pathology & Microbiology	BTP 121, BTP 221, BTP 321, BTP/BTR 324.
5.	<b>Recombination Techniques</b>	BTR 211, BTI/BTR 221, BTR 311, BTI/BTR 321,
		BTR 322, BTP/BTR 324, BTI/BTR 328, BTI/BTR
		329.
6.	Finger Printing	BTF 311, BTF 312, BTF 321.
7.	Immunology & Defense	BTI/BTR 221, BTI/VPB/VMC 312, BTI 311,
''	Mechanism	BTI/BTR 321, BTI/VPB/VMC 323, BTI/VPB 324,
		BTI/VPB 325, BTI/VMC 326, BTI/VPT 327,
		BTI/BTR 328, BTI/BTR 329, BTI/VPT 330.
8.	<b>Commercial Biotechnology</b>	BTO 111, BTO 121, BTO/AGE 321, BTO 322,
		BTO/AGE 411
9.	College Level	NCC/NSS/PEY 111, BTS 411, BTS 412, BTS 421,
		BTS 422.

# Department wise distribution of courses

### College of Biotechnology

	Conege of Biotechnology		
Cell Biology	Course	Credit	Page
BTC 111	Cell Biology	2 (2-0-0)	20
BTC 121	Biodiversity and its conservation	2 (2-0-0)	27
BTC 221	Classical and Molecular Cytogenetics	3 (2-0-1)	41
BTC 321	Molecular Ecology and Evolution	3 (3-0-0)	60
DIC 521	Wolceular Leology and Evolution	5 (5-0-0)	00
Dhusiology & Dioch	and stars		
Physiology & Bioch		2(2,0,1)	26
BTB 211	Plant Physiology	3 (2-0-1)	36
BTB/ AGE 221	Electronics and Instrumentation in Biotechnology	2 (1-0-1)	38
BTB 222	General Biochemistry	4 (3-0-1)	39
BTB 311	Enzymology & Enzyme Technologies	3 (2-0-1)	48
BTB/BAS 327	Metabolomics and System Biology	3 (2-0-1)	66
	& Genetic Engineering		
BTM 111	Basic Genetics	3 (2-0-1)C*	19
BTM 121	Molecular Biology	3 (2-0-1)	26
BTM 221	Plant Genetic Transformation	3 (2-0-1)	41
BTM 311	Molecular Genetics	2 (2-0-0)	45
BTM 312	Nanobiotechnology	2 (2-0-0)	46
BTM 321	Principles and Applications of Plant	. ,	
	Genetic Transformation	3 (2-0-1)	51
BTM 322	Epigenetics and Gene Regulation	3 (2-0-1)	52
		- ()	• –
Pathology & Microl	biology		
BTP 121	Microbiology	3 (2-0-1)	28
BTP 221	Microbial Genetics	3 (2-0-1)	42
BTP 321	Microbial Biotechnology	3 (2-0-1)	59
BTP/ BTR 324	Green Biotechnology	3 (2-0-1)	60
DIII DIR 521	Sicci Diotechiology	5 (2 0 1)	00
<b>Recombination Tecl</b>	hniques		
BTR 211	Recombinant DNA Technology	3 (2-0-1)	35
BTI/ BTR 221	Introductory Bioinformatics	3 (2-0-1)	40
BTR 311	Genomics and Proteomics	3 (3-0-0)	47
BTI/ BTR 321	Computational Biology	3 (2-0-1)	49
BTR 322	Applications of Genomics and Proteomics	3 (2-0-1)	52
BTP/ BTR 324	Green Biotechnology	3 (2-0-1)	60
		· /	
BTI/ BTR 328	Bioinformatics Tools and Biological Databases	3(2-0-1)	64
BTI/ BTR 329	Structural Bioinformatics	3 (2-0-1)	65
Finaar Drinting			
Finger Printing	Malagular Markor Tashrala av	2(200)	16
BTF 311	Molecular Marker Technology	2 (2-0-0)C	46
BTF 312	IPR, Biosafety and Bioethics	2(2-0-0)	47
BTF 321	Plant Tissue Culture and its Applications	3 (2-0-1)	50

### Immunology & Defense Mechanism

BTI/ BTR 221	Introductory Bioinformatics	3 (2-0-1)	40
BTI/ VPB/VMC 312	65	4 (3-0-1)	44
BTI 311	Immunology	3 (2-0-1)	45
BTI/BTR 321	Computational Biology	3 (2-0-1)	49
BTI/VPB/VMC 323	Principles and Procedures of Animal Cell Culture	3 (2-0-1)	56
BTI/VPB 324	Transgenic Animal Production	3 (3-0-0)	57
BTI/VPB 325	Molecular Diagnostics	3 (2-0-1)	57
BTI/VMC 326	Molecular Virology and Vaccine Production	3 (2-0-1)	58
BTI/VPT 327	Molecular Pharming and Biopharmaceuticals	3 (2-0-1)	61
BTI/ BTR 328	Bioinformatics Tools and Biological Databases	3 (2-0-1)	64
BTI/ BTR 329	Structural Bioinformatics	3 (2-0-1)	65
BTI/ VPT 330	Pharmacogenomics	3 (2-0-1)	65
	-		
<b>Commercial Biotech</b>	inology		
BTO 111	Introduction to Biotechnology	3 (2-0-1)	20
BTO 121	Plant Tissue Culture	3 (2-0-1)	26
BTO/AGE 321	Food Biotechnology	3 (2-0-1)	61
BTO 322	Bio-prospecting of Molecules and Genes	3 (3-0-0)	62
BTO/AGE 411	Bioprocess Engineering, Bioseperation and		
	Downstream processing	3 (3-1-0)C	68
	1 0		
<b>Biotechnology</b> Colleg	ge Level		
	NCC/NSS/PEY (NG)	2 (0-2-0)	XX
BTS 411	Educational Tour (NG)	1 (0-1-0)	67
BTS 412	Student READY – (04 Optional topics)		
	In-house Skill Development Modules	20 (0-0-20)	67
BTS 421	Student READY - Project Formulation,		
	Execution and Presentation	10	69
BTS 422	Student READY - Entrepreneurial Development		
	in Biotechnology	10	69
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### **College of Agriculture**

<b>Basic Science</b>	0 0		
BAS 112	Human Ethics	1 (1-0-0)	13
BAS 116	Basic Mathematics-I	3 (3-0-0)	13
BAS 114	Basic Botany	3 (2-0-1)	14
BAS 115	Communication Skills & Personality Development		15
BAS 121	Basic Zoology	3 (2-0-1)	21
BAS 122	Basic Mathematics- II	3 (3-0-0)	21
BAS 123	Basic Statistics	2 (1-0-1)	22
BAS 211	Biomathematics	3 (2-0-1)	29
BAS 214	Information and Communication Technology	2 (1-0-1)	29
BAS 221	Biophysics	3 (2-0-1)	36
BAS 311	Agricultural Informatics	3 (2-0-1)	43
BAS 321	Biostatistics	3 (2-0-1)	49
BAS 322	Programming for Bioinformatics	4 (2-0-2)	63
BAS 323	Computational Methods for Data Analysis	2 (1-0-1)	63
BTB/BAS 327	Metabolomics and System Biology	3 (2-0-1)	66
Agriculture Enginee	ring & Food Technology		
AGE/ SAC/	Environmental Studies and Disaster Management	3 (2-0-1)C	15
AGR 114	č	× ,	
AGE 113	Food Science and Processing	2 (1-0-1)	17
BTB/ AGE 221	Electronics and Instrumentation in Biotechnology	2 (1-0-1)	38
BTO/AGE 321	Food Biotechnology	3 (2-0-1)	61
BTO/AGE 411	Bioprocess Engineering, Bioseperation and	× ,	
	Downstream processing	3 (3-1-0)	68
Agronomy		~ /	
AGE/ SAC/	Environmental Studies and Disaster Management	3 (2-0-1)C	15
AGR 114	C		
AGR 113	Crop Production Technology	3 (2-0-1)C	18
Soil Science and Agr	riculture Chemistry		
AGE/ SAC/	Environmental Studies and Disaster Management	3 (2-0-1) C	15
AGR 114			
Genetics and Plant H	Breeding		
GPB 122	Basics of Plant Breeding	3 (2-0-1)	24
GPB 212	Breeding of Field Crops	3 (2-0-1)	33
GPB 322	Molecular Breeding in Field Crops	3 (2-0-1)	53
Entomology			
ENT/ PPA 212	Fundamentals of Crop Protection	3 (2-0-1)	31
Plant Pathology			
ENT/ PPA 212	Fundamentals of Crop Protection	3 (2-0-1)	31

	TT /• 1/			
Horticulture HOR 121 HOR 322		Production Technologies for Horticultural Crops Molecular Breeding of Horticultural Crops	3 (2-0-1)	23
		and Forest Trees	3 (2-0-1)	54
	Agricultural Econor	nics		
	AAE 212	Economics and Marketing	3 (2-0-1)	30
	AEC/ AAE 222	Entrepreneurship Development and Business	0 (1 0 1)	
		Management	2 (1-0-1)	37
	Agriculture Extension			
	AEC/ AAE 222	Entrepreneurship Development and Business	<b>2</b> (1, 0, 1)	27
	Animal Production	Management	2 (1-0-1)	37
	AAP 121	Introduction to Animal Breeding	3 (2-0-1)C	25
		<b>College of Veterinary Sciences</b>		
	Veterinary Physiolo	gy and Biochemistry		
	VAN/VPB 123	Anatomy and Physiology of Livestock	3 (3-0-0)	24
	BTI/VPB/VMC 312	Animal Biotechnology	4 (3-0-1)	44
	VPB 321	Animal Genomics	3 (2-0-1)	55
	VPB/VGO 322	Embryo Transfer Technologies	3 (2-0-1)	55
	BTI/VPB/VMC 323	Principles and Procedures of Animal Cell Culture	3 (2-0-1)	56
	BTI/VPB 324	Transgenic Animal Production	3(3-0-0) 2(2,0,1)	57
	BTI/VPB 325	Molecular Diagnostics	3 (2-0-1)	57
	Livestock Product T			~ -
	LPT 211	Livestock Product Technology	3 (2-0-1)	35
	Veterinary Microbio	ology		
	VMD/VMC 211	Animal Health Care	3 (2-0-1)C	34
	BTI/VPB/VMC 312	65	4 (3-0-1)	44
	BTI/VPB/VMC 323	Principles and Procedures of Animal Cell Culture	3 (2-0-1)	56
	BTI/VMC 326	Molecular Virology and Vaccine Production	3 (2-0-1)	58
	Livestock Productio			
	LPM 212	Livestock Production and Management	3 (2-0-1)	34
	Veterinary Medicine	e		
	VMD/VMC 212	Animal Health Care	3 (2-0-1)C	34
	Veterinary Anatomy			
	VAN/VPB 123	Anatomy and Physiology of Livestock	3 (3-0-0)	24
Veterinary Gynecology & Obstetrics				
	VPB/VGO 322	Embryo Transfer Technologies	3 (2-0-1)	55
Veterinary Pharmacology & Toxicology				
	BTI/ VPT 330	Pharmacogenomics	3 (2-0-1)	65
	BTI/VPT 327	Molecular Pharming and Biopharmaceuticals	3 (2-0-1)	61

#### **SEMESTER-I**

<b>Course Title</b>	:	Human Ethics
Course No.	:	BAS 112
<b>Credit Hours</b>	:	1 (1-0-0)

#### <u>UNIT I</u>

Universal human aspirations: Happiness and prosperity; Human values and ethics: Concept, definition, significance and sources; Fundamental values: Right conduct, peace, truth, love and non-violence; Ethics: professional, environmental, ICT; Sensitization towards others particularly senior citizens, developmentally challenged and gender.

#### <u>UNIT II</u>

Spirituality, positive attitude and scientific temper; Team work and volunteering; Rights and responsibilities; Road safety; Human relations and family harmony; Modern challenges and value conflict: Sensitization against drug abuse and other social evils; Developing personal code of conduct (SWOT Analysis); Management of anger and stress.

#### **Suggested Readings**

Gaur RR, Sangal R & Bagaria GP. 2011. A Foundation Course in Human Values and Professional Ethics. Excel Books.

- Mathur SS. 2010. Education for Values, Environment and Human Rights. RSA International.
- Sharma RA. 2011. Human Values and Education -Axiology, Inculcation and Research. R. Lall Book Depot.
- Sharma RP & Sharma M. 2011. Value Education and Professional Ethics. Kanishka Publishers.

Srivastava S. 2011. Human Values and Professional Ethics. S K Kataria & Sons.

Srivastava S. 2011. Environmental Science. S K Kataria & Sons.

Tripathi A.N. 2009. Human Values. New Age International (P) Ltd Publishers.

<b>Course Title</b>	:	<b>Basic Mathematics-I</b>
Course No.	:	BAS 116
<b>Credit Hours</b>	:	3 (3-0-0)

#### <u>UNIT I</u>

Complex numbers: Properties of real numbers, complex numbers, their addition, multiplication and division, square root of complex numbers, cube roots of unity and their properties, De-Moivre's theorem; Theory of equations: Solution of quadratic equation, equation reducible to quadratic equation, relation between roots and coefficients, nature of roots and formation of quadratic equation with given roots.

#### <u>UNIT II</u>

Geometric series: nth term of G.P. series, sum of G.P. series, geometric mean; Harmonic

series, harmonic mean; Arithmetico geometric series and special series  $\sum n$  ,

 $\sum n^2$ ,  $\sum n^3$ . Partial fractions; Logarithms; Binomial theorem for any index: Expansion, middle term, general term, terms independent of x. <u>Unit III</u>

Trigonometry: Trigonometric ratios, allied angles, graphs of trigonometric functions; Addition and subtraction formulae; Product and sum formulae; Multiple and sub-multiple angles, sine, cosine and projection formulae; Area of a triangle.

#### **Suggested Readings**

NCERT 2012. *Mathematics of Class XI*. NCERT India. Sharma RD. 2014. *Mathematics of Class XI*. Dhanpat Rai Publisher.

<b>Course Title</b>	:	<b>Basic Botany</b>
Course No.	:	BAS 114
<b>Credit Hours</b>	:	3 (2-0-1)

#### <u>UNIT I</u>

Plant kingdom and features of each group; Morphology, modifications and functions of root, stem, leaf, flower and inflorescence; Pollination and fertilization; Fruit types; Structure of dicot and monocot seed, seed germination.

#### <u>UNIT II</u>

Cell structure; DNA, chromosome and genes; Cell and tissue types; Internal structure of root, stem and leaf.

#### <u>UNIT III</u>

Plant taxonomy, systems of classification; Characteristics and economic importance of Poaceae, Brassicaceae, Fabaceae, Malvaceae, Rutaceae, Rosaceae, Asteraceae and Solanaceae families

#### Practical

Description of one plant species from each group of plant kingdom; Study of morphology and modifications of root, stem, leaf, flower; Types of inflorescence; Structure of various types of seeds and fruits; Demonstration of cell structure, tissue types; Structure of monocot and dicot root, stem and leaf; One flower from each family.

#### **Suggested Readings**

Bendre A & Kumar A. 1999. *Textbook of Practical Botany*. Vol. 2, 7th Ed., Rastogi Publications.

Bendre AM & Pande PC. 2009. *Introduction to Botany*. Rastogi publications. Dutta AC. 1995. *A Class Book of Botany*, 16th Edition. Oxford University Press.

<b>Course Title</b>	:	<b>Communication Skill and Personality Development</b>
Course No.	:	BAS 115
<b>Credit Hours</b>	:	2 (1-0-1)

#### <u>UNIT I</u>

Communication skills: Structural and functional grammar; Meaning and process of communication; Verbal and nonverbal communication; Listening and note taking; Writing skills; Oral presentation skills; Field diary and lab record; Indexing, footnote and bibliographic procedures; Reading and comprehension of general and technical articles; Precise writing, summarizing, abstracting; Individual and group presentations; Impromptu presentation; Public speaking; Group discussion and interviews; Organizing seminars and conferences.

#### <u>UNIT II</u>

Voice modulation basics and their usage for meaningful impact on people; Attributes of an effective leader; Stress and conflict management; Time management: Personal organization, prioritizing and balancing; Cosmopolitan culture; Impact of non verbal communication; Science of body language; Role of team work.

#### Practical

Listening and note taking, writing skills, oral presentation skills; Field diary and lab record; Indexing, footnote and bibliographic procedures; Reading and comprehension of general and technical articles, precise writing, summarizing, abstracting; Individual and group presentations; Mock group discussions and interviews; Attitude management; Setting and achieving a short term goal; Creating a personal vision statement of life; Voice modulation; Practicing conscious body postures and movements; Rapport building; Team work exercises; Time management.

#### **Suggested Readings**

Carnegie, Dale. 2012. How to Win Friends and Influence People in the Digital Age. Simon & Schuster.

Covey Stephen R. 1989. TheSeven Habits of Highly Successful People. Free Press.

Spitzberg B, Barge K & Morreale, Sherwyn P. 2006. Human Communication: Motivation, Knowledge & Skills.Wadsworth.

Verma, KC. 2013. The Art of Communication. Kalpaz.

<b>Course Title</b>	:	<b>Environmental Studies and Disaster Management</b>
Course No.	:	AGE/SAC/AGR 114
<b>Credit Hours</b>	:	3 (2-0-1)

#### ENVIRONMENTAL STUDIES

<u>UNIT I</u>

Multidisciplinary nature of environmental studies; Definition, scope and importance.

#### <u>UNIT II</u>

Natural Resources: Renewable and non-renewable resources; Natural resources and associated problems.

a) Forest resources: Use and over-exploitation; Deforestation; Case studies. Timber extraction, mining; Dams and their effects on forest and tribal people.

b) Water resources: Use and over-utilization of surface and ground water; Floods; Drought; Conflicts over water; Dams-benefits and problems.

c) Mineral resources: Use and exploitation; Environmental effects of extracting and using mineral resources; Case studies.

d) Food resources: World food problems; Changes caused by agriculture and overgrazing; Effects of modern agriculture; Fertilizer-pesticide problems; Water logging; Salinity; Case studies.

e) Energy resources: Growing energy needs; Renewable and non-renewable energy sources; Use of alternate energy sources; Case studies.

f) Land resources: Land as a resource; Land degradation; Man induced landslides; Soil erosion and desertification.

Role of an individual in conservation of natural resources; Equitable use of resources for sustainable lifestyles.

#### <u>UNIT III</u>

Ecosystems; Concept of an ecosystem; Structure and function of ecosystem; Producers, consumers and decomposers; Energy flow in ecosystem; Ecological succession; Food chains, food webs and ecological pyramids; Introduction, types, characteristic features, structure and function of forest ecosystem, grassland ecosystem, desert ecosystem and aquatic ecosystems (ponds, streams, lakes, rivers, oceans, estuaries)

#### <u>UNIT IV</u>

Biodiversity and its conservation; Introduction, definition, genetic, species and ecosystem diversity and biogeographical classification of India; Value of biodiversity: consumptive use, productive use, social, ethical, aesthetic and option values; Biodiversity at global, national and local levels; India as a mega-diversity nation; Hot-sports of biodiversity; Threats to biodiversity: habitat loss, poaching of wildlife, man-wildlife conflicts; Endangered and endemic species of India; Conservation of biodiversity: *In-situ* and *Exsitu* conservation of biodiversity.

#### UNIT V

Environmental Pollution: definition, cause, effects and control measures air pollution, water pollution, soil pollution, marine pollution, noise pollution, thermal pollution, nuclear hazards; Solid waste management: causes, effects and control measures of urban and industrial wastes; Role of an individual in prevention of pollution; Pollution case studies.

#### <u>UNIT VI</u>

Social issues and the environment; From unsustainable to sustainable development; Urban problems related to energy; Water conservation, rain water harvesting, watershed management; Environmental ethics: Issues and possible solutions, climate change, global warming, acid rain, ozone layer depletion, nuclear accidents and holocaust; Wasteland reclamation; Consumerism and waste products; Environment Protection Act; Air

(Prevention and Control of Pollution) Act; Water (Prevention and control of Pollution) Act; Wildlife Protection Act; Forest Conservation Act; Issues involved in enforcement of environmental legislation; Public awareness.

#### DISASTER MANAGEMENT

#### UNIT I

Natural disasters - Meaning and nature of natural disasters; their types and effects. Floods, drought, cyclone, earthquakes, landslides, avalanches, volcanic eruptions, heat and cold waves; Climatic change: global warming, sea level rise, ozone depletion.

#### UNIT II

Man-made disasters - Nuclear disasters, chemical disasters, biological disasters, building fire, coal fire, forest fire, oil fire, air pollution, water pollution, deforestation, industrial waste water pollution, road accidents, rail accidents, air accidents, sea accidents.

#### UNIT III

Disaster management - Effect to migrate natural disaster at national and global levels; International strategy for disaster reduction; Concept of disaster management; National disaster management framework; Financial arrangements; Role of NGOs, communitybased organizations and media; Role of central, state, district and local administration; Armed forces, police and other organizations in disaster response.

#### Practical

Visit to a local area to document environmental assets: river/ forest/ grassland/ hill/ mountain; Visit to a local polluted site - urban/ rural/ industrial/ agricultural; Study of common plants, insects, birds and study of simple ecosystems - pond, river, hill slopes, etc.; Visit to disaster management organizations; Collection of statistics of national disasters occurred since 20<sup>th</sup> century.

#### **Suggested Readings**

Ahluwalia VK & Malhotra S. 2006. Environmental Science. Ane Books India.

Anjaneyulu Y. 2004. Introduction to Environmental Science. BS Publications. Chauhan AS. 2009. Environmental Studies. 3rd Edition. Jain Brothers.

Das RC & Behera DK. 2008. Environmental Science - Principles and Practice. Prentice-Hall of India Pvt Ltd.

Dhaliwal GS & Kukal SS. 2005. Essentials of Environment Science. Kalyani Publishers. Santra

<b>Course Title</b>	:	Food Science and Processing
Course No.	:	AGE 113
<b>Credit Hours</b>	:	2 (1-0-1)

#### <u>UNIT I</u>

Definition: Food and nutrition; Food production and consumption trends in India; Major deficiencies of calories, proteins, vitamins and micronutrients; Food groups and concept of balanced diet: RDA.

#### <u>UNIT II</u>

Causes of food spoilage; Principles of processing and preservation of food by heat, low temperature, drying and dehydration, chemicals and fermentation; Preservation through ultraviolet and ionizing radiations.

#### <u>UNIT III</u>

Post-harvest handling and technology of fruits, vegetables, cereals, oilseeds, milk, meat and poultry; Food safety, adulteration and food laws; Status of food industry in India.

#### Practical

Physical and chemical quality assessment of cereals, fruits, vegetables, egg, meat and poultry; Value added products from cereals, millets, fruits, vegetables, milk, egg and meat; Visit to local processing units.

#### **Suggested Readings**

Potter NN & Hotchkiss JH. 1995. *Food Science*. Chapman and Hall Publishers. Swaminathan M. 2005. *Handbook of Foods and Nutrition*. Ganesh and Co. Pvt. Ltd. Swaminathan M. 1990. *Food Science, Chemistry and Experimental Foods*. BAPPCO. Vickie A., Vaclavik& Elizabeth W. Christian. 2003. Essentials of Food Science, 2<sup>nd</sup>Ed.

Kluwer Academic/ Plenum Publishers, New York.

<b>Course Title</b>	:	<b>Crop Production Technology</b>
Course No.	:	AGR 113
<b>Credit Hours</b>	:	3 (2-0-1)

#### <u>UNIT I</u>

Soil and its components; Soil morphological, physical, chemical and biological properties; Acidic, saline and alkali soils and their reclamation; Essential plant nutrients: Functions and deficiency symptoms; Soil micro-organisms; Rhizosphere and its domain in soil; Organic manures and inorganic fertilizers.

#### <u>UNIT II</u>

Agriculture; Agronomy and its relation with other sciences; Classification of crops; Tillage and tillage practices, concepts of tillage and objectives; Seed, its characteristics and different sowing methods; Weed management: definition of weed, losses and benefits of weeds, different weed control methods and their suitability under different conditions; Irrigation: Soil water classification, methods of irrigation, approaches for scheduling irrigation.

#### <u>UNIT III</u>

Soil fertility and productivity; Concept of essentiality of plant nutrients; Fertilizers, manures and their types, methods of fertilizer application; Concepts of crop rotation, multiple cropping and intercropping - their principles, advantages and limitations; Cropping intensity; Production technology of major crops: Rice, maize, cotton, soybean, mung bean, mash, wheat, rapeseed and mustard, gram and Egyptian clover.

#### Practical

Study of soil profile and its characteristics; Determination of soil particle size distribution, particle density and bulk density; Determination of soil pH, electrical conductivity and organic carbon; Study of soil micro-flora (bacteria, fungus and actinomycetes).

Land measurement; Practice in seedbed preparation and seeding methods; Identification of crop seeds, crops, weeds and fertilizers; Identification and use of hand tools and implements; Computation of fertilizer doses and their method of application.

#### **Suggested Readings**

Acquaah G. 2005. Principles of Crop Production: Theory, Techniques and Technology. Prenice Hall.

Alexander M. 1977. Introduction to Soil Microbiology, 2<sup>nd</sup> Edition. John Wiley & Sons.

Balasubrananiyan P & Palaniappan SP. 2010. Principles and Practices of Agronomy. Agrobios.

Brady NC & Well RR. 2002. The Nature and Properties of Soils, Thirteenth Edition. Pearson Prentice Hall.

Chandrasekaran B, Annadural K & Samasundaram E. 2010. A Text Book of Agronomy. New Age International (P) Limited Publishers.

Das DK. 2011. Introductory Soil Science. Third Revised Edition, Kalyani Publishers. Reddy SR. 2011. Principles of Agronomy. Kalyani Publishers.

<b>Course Title</b>	:	<b>Basic Genetics</b>
Course No.	:	BTM 111
<b>Credit Hours</b>	:	3 (2-0-1)

#### <u>UNIT I</u>

History of Genetics; Mendel's principles and rediscovery; Cell division; Chromosomes structure and function; Chromosome theory of inheritance; Sex-linked, sex-limited and sex-influenced inheritance; Sex determination and sex differentiation.

#### <u>UNIT II</u>

Multiple allelism; Linkage and crossing-over; Gene-gene interaction; Genetic analysis in prokaryotes and eukaryotes; Extra chromosomal inheritance; Mutations; Hardy-Weinberg law; Quantitative inheritance; Introduction to Human genetics; Genetic basis of evolution.

#### Practical

Life cycle in model plants and animals; microscopy; Mitosis and meiosis; Monohybrid crosses (segregation); Dihybrid crosses (independent assortment); Probability and use of Chi-square; Sex-linked inheritance; Multiple allelism; Detection and estimation of linkage. (\*to be offered by Agriculture College).

#### **Suggested Readings**

Gupta PK. 2014. *Genetics* 4th ed. Rastogi Publications. Inbasekar P. 2009. *Cell Biology and Genetics*. Panima Publications. Miglani GS. 2000. Basic Genetics. Narosa Publishing house, New Delhi.

Russell PJ. 2013. *iGenetics: Pearson New International Edition: A Molecular Approach*. Pearson.

Watson JD, Bakee TA, Bell SP, Gann A, Levine M & Losick R. 2008. *Molecular Biology of the Gene*. 6th Ed. Pearson Education International.

<b>Course Title</b>	:	Introduction to Biotechnology
Course No.	:	BTO 111
<b>Credit Hours</b>	:	3 (2-0-1)

#### <u>UNIT I</u>

History, definitions, concepts, scope and importance of Biotechnology: Plant, microbial, animal, medical, environmental, industrial, Marine, Agricultural and food Biotechnology; Nanobiotechnology.

#### <u>UNIT II</u>

Introduction to recombinant DNA technology and its applications: Vectors, DNA restriction and modifying enzymes, gene cloning; Introduction to genomics and proteomics: Molecular markers, DNA sequencing; Genetic transformation and transgenic organisms; Bioinformatics. Biosafety guidelines.

#### Practical

Orientation to the laboratories: glass houses, screen houses, transgenic facilities and field area; General guidelines for working in Biotechnology laboratories; Familiarization with basic equipment's used in biotechnology; Selection of chemicals (different grade), buffer preparation, calculations and scientific notations used in laboratories.

#### Suggested Readings

Brown T A. 2002. Genomes 2. 2nd ed. New

york:Wiley-Liss.

- Prave P, Faust U, Sittig W & Sukatsch DA. 1987. *Basic Biotechnology: A Student's Guide*. VCH Verlagsgesellschaft.
- Prave P, Faust U & Sittig W. 1987. *Fundamentals of Biotechnology*. VCH Verlagsgesellschaft.

Renneberg R. 2008. Biotechnology for Beginners. Academic Press Publishers.

<b>Course Title</b>	:	Cell Biology
Course No.	:	BTC 111
<b>Credit Hours</b>	:	2 (2-0-0)

#### <u>UNIT I</u>

Origin and evolution of cell; Introduction to microscopy; Sub-cellular structure of prokaryotic and eukaryotic cells; Membrane structure and function: plasma membrane, cell wall and extracellular matrix; Structural organization and function of intracellular organelles and organelle biogenesis: Nucleus, mitochondria, endoplasmic reticulum, Golgi apparatus, lysosomes, peroxisomes, plastids, vacuoles.

#### <u>UNIT II</u>

Structure and function of cytoskeleton and its role in motility; Cell membrane transport; Introduction to cell signalling; Cell growth, cell cycle and its control; Cell death and cell renewal.

#### **Suggested Readings**

Alberts B, Johnson A, Lewis J, Raff M, Roberts K &Walter P. 2008. *Molecular Biology* of the Cell. 5<sup>th</sup> Ed. Garland Science/ Taylor and Francis Group.

Lodish H, Berk A, Kaiser CA, Krieger M, Bretscher A, Ploegh H, Amon A & Scott MP. 2012. *Molecular Cell Biology*. W. H. Freeman.

Sadava DE. 1993. *Cell Biology: Organelle Structure and Function*. Jones and Bartlett Publishers.

#### **SEMESTER-II**

<b>Course Title</b>	:	<b>Basic Zoology</b>
Course No.	:	BAS 121
<b>Credit Hours</b>	:	3 (2-0-1)

#### <u>UNIT I</u>

Introduction to Zoology; Structure and functions of cell and cell organelles; Difference between prokaryotic and eukaryotic cell; Cell division – mitosis and meiosis; Structure and function of biomolecules; Types of simple and compound tissues.

#### <u>UNIT II</u>

Binomial Nomenclature; Classification and general survey of animal kingdom; Functional organization of various systems of a mammal: digestive, circulatory, respiratory, excretory, nervous and reproductive; Laws of inheritance; Multiple allelism blood groups; Genetic disorders in human and their inheritance.

#### Practical

Study of animal cell structure and cell division; Histological preparation of simple and compound tissues; General survey of animal kingdom up to phyla in invertebrates and up to classes in vertebrates; Demonstration of mammalian anatomy; Blood grouping.

#### **Suggested Readings**

Bhatia KN & Tyagi MP. 2012. *Trueman's Elementary Biology*. 24th ed. Trueman Book Company.

Dhami PS & Mahindru RC. 1996. *A Text Book of Biology for 10+2*. Pradeep Publications.

<b>Course Title</b>	:	<b>Basic Mathematics-II</b>
Course No.	:	BAS 122
<b>Credit Hours</b>	:	3 (3-0-0)

#### <u>UNIT I</u>

Functions; Limit: Introduction, left handed and right handed limits, general rules for

calculation of limits Standard limits  $\underset{x\to 0}{\text{Lt}} \frac{x^n - a^n}{x - a}$ ,  $\underset{x\to 0}{\text{Lt}} \frac{\sin x}{x}, \underset{x\to 0}{\text{Lt}} (1 + x)^{\frac{1}{x}}, \underset{x\to 0}{\text{Lt}} \frac{a^x - 1}{x}, \underset{x\to 0}{\text{Lt}} \frac{\log(1 + x)}{x}$ . Continuity: Definition of

continuity, continuity of algebraic functions, Continuity of trigonometric and exponential functions.

#### <u>UNIT II</u>

Differentiation: Differentiation by first principle, sum, difference, product and quotient formulae, differentiation using chain rule, differentiation of functions in parametric and implicit form, logarithmic differentiation, geometrical interpretation of derivative, Successive differentiation, geometrical interpretation of derivative, maxima and minima, tangent and normal.

#### <u>UNIT III</u>

Integration: Integration by substitution, integration by partial fractions, integration by parts, integration by trigonometric substitution.

#### <u>UNIT IV</u>

Matrices and Determinants: Definition of matrix, addition, subtraction and multiplication, inverse of matrix; Solution of linear equations: By Crammer's rule and inverse of matrix.

#### **Suggested Readings**

NCERT 2012. *Mathematics of Class XII*. NCERT India. Sharma RD. 2014. *Mathematics of Class XII*. Dhanpat Rai Publisher.

<b>Course Title</b>	:	<b>Basic Statistics</b>
Course No.	:	<b>BAS 123</b>
<b>Credit Hours</b>	:	2 (1-0-1)

#### <u>UNIT I</u>

Definition of statistics, its use and limitations; Frequency distribution and frequency curve and cumulative frequency curve; Measures of central tendency; Measures of dispersion; Probability: Definition, additive and multiplicative law for two events; Normal distribution and its properties; Introduction to sampling; Sampling techniques.

#### UNIT II

Tests of significance: Null hypothesis, alternate hypothesis, Type I & II Error, one and two tail tests, level of significance and confidence interval; SND test for means: Single sample and two samples Z-test; Student's t-test for means, single sample, two samples and paired t-test; F-test;

#### UNIT III

Chi-square test in 2x2 contingency table; Yate's correction for continuity; Correlation: Scatter diagram and Karl Pearson's coefficient of correlation for ungrouped data and its

testing; Linear regression and its properties; Analysis of variance and its assumptions, Analysis of CRD and RBD; Analysis of Latin Square Design.

#### Practical

Construction of frequency distribution tables and frequency curves; Computation of Arithmetic: Mean, median, mode; Standard deviation; Variance and coefficient of variation for ungrouped and grouped data; SND test for means; Student's t-test; F-test and Chi-square test; Correlation coefficient 'r' and its testing; Fitting of regression equations; Analysis of CRD, RBD and LSD.

#### **Suggested Readings**

Freud JE & Perles BM. 2006. *Modern Elementary Statistics*. 12<sup>th</sup> Ed. Pearson India. Kapoor VK. 2003. *Problems and Solutions in Statistics*. 7<sup>th</sup> Edition. Sultan Chand and Sons.

Snedecor GW. & Cochran WG. 1989. Statistical Methods. Iowa State University Press.

<b>Course Title</b>	:	<b>Production Technologies for Horticultural Crops</b>
Course No.	:	HOR 121
<b>Credit Hours</b>	:	3 (2-0-1)

#### <u>UNIT I</u>

Importance and scope of fruit cultivation; Classification of fruit crops; Climatic requirement; Selection of site; Fencing and wind break; Lay out and planting systems; Sexual and asexual methods of plant propagation; Production technology of important tropical, sub tropical and temperate fruit crops.

#### <u>UNIT II</u>

Importance of vegetable cultivation for nutritional security; Production technology of important vegetable crops: potato, brinjal, tomato, chilli, onion, okra, cabbage, cauliflower, musk melon, water melon, cucumber and leafy vegetables.

#### <u>UNIT III</u>

Status and scope of floriculture in India and abroad; Production technology of commercial flower crops: Rose, chrysanthemum, gladiolus, marigold, gerbera, carnation, lilium, jasmine, anthurium and orchids.

#### Practical

Identification of different fruit, vegetables, ornamental and flower crops; Lay out and planning for planting orchards; Preparation of seed beds; Raising of seeds, rootstocks, and propagation techniques of major fruit, vegetable and flower crops; Visit to commercial nurseries and orchards.

#### **Suggested Readings**

Arora JS. 2013. Introductory Ornamental Horticulture. Kalyani Publishers. Bal JS. 2013. Fruit Growing. Kalyani Publishers. Chadha KL. 2012. Handbook of Horticulture. ICAR Dhaliwal MS. 2014. Handbook of Vegetable Crops. Kalyani Publishers

<b>Course Title</b>	:	Anatomy and Physiology of Livestock
Course No.	:	VAN/VPB 123
<b>Credit Hours</b>	:	3 (3-0-0)

#### UNIT I

Definition of terms used in veterinary anatomy, topography, contour, landmarks and functional anatomy of various organs in cow, buffalo, sheep and goat structural and functional classification of muscles.

#### <u>UNIT II</u>

Structure of animal cell and tissues: study of microscopic structure of organs from digestive, urinary, respiratory, reproductive, nervous, cardiovascular and endocrine systems; Gametogenesis, fertilization, cleavage, gastrulation and the development of fetal membranes in livestock, structure and types of mammalian placenta; Development of the organs of digestive, urogenital, cardiovascular, nervous and endocrine glands.

#### <u>UNIT III</u>

Introduction to blood physiology; Genetic and endocrine control of reproductive system; maternal recognition of pregnancy; Introduction to physiology of mammary glands: structure and development, hormonal control of mammary growth, lactogenesis and lactation cycle.

#### **Suggested Readings**

Keith Dyce & Wolfgang Sack & Wensing CJG. 2009. *Textbook of Veterinary Anatomy*, 4th Edition. Elsevier Health Sciences.

Rowen D. Frandson, W. Lee Wilke & Anna Dee Fails. 2009. *Anatomy and Physiology of Farm Animals*. 7<sup>th</sup> Edition. Wiley–Blackwell.

William O, Reece. 2004. *Duke's Physiology of Domestic Animals*. 12<sup>th</sup> Edition. Comstock Publishing Associates.

<b>Course Title</b>	:	<b>Basics of Plant Breeding</b>
Course No.	:	GPB 122
<b>Credit Hours</b>	:	3 (2-0-1)

#### <u>UNIT I</u>

History, aims and objectives of Plant breeding; Role of related sciences in plant breeding; Modes of reproduction - sexual, asexual, apomixes: Significance in plant breeding; Modes of pollination, genetic consequences, differences between self- and cross pollinated crops; Germplasm resources and their utilization.

#### <u>UNIT II</u>

Methods of breeding: Introduction and Acclimatization; Selection: Mass selection, Johannesen's pure-line theory, genetic basis, pure-line selection; Hybridization: Aims and objectives, types of hybridization; Methods of handling segregating generations: Pedigree method, bulk method, back cross method; Heterosis, inbreeding depression, various theories of heterosis, exploitation of hybrid vigor, Hardy Weinberg law, selection

in cross pollinated crops; Population improvement programmes; Synthetics and composites; Methods of breeding vegetatively propagated crops.

#### <u>UNIT III</u>

Incompatibility and male sterility and their utilization in crop improvement; Mutation breeding; Ploidy breeding; Wide hybridization and its significance in crop improvement; Procedure for release of new varieties.

#### Practical

Classification of plants; Botanical description and floral biology of field crops: rice, sorghum, maize, wheat, bajra, sugarcane, brassicas, groundnut, sunflower, sesamum, red gram, bengal gram, green gram, soybean, black gram, cotton; Study of megasporogenesis and microsporogenesis; Fertilization and life cycle of an angiospermic plant; Hybridization techniques and precautions to be taken; selfing, emasculation and crossing techniques; Study of male sterility and incompatibility.

#### **Suggested Readings**

Allard RW. 1960. Principles of Plant Breeding. John Wiley and Sons.
Chahal GS & Gosal SS. 2002. Principles and Procedures of Plant Breeding: Biotechnological and Conventional Approaches. Narosa Publishers.

Phundan Singh. 2014. Essentials of Plant Breeding. Kalyani Publishers. Singh BD. 2009. Plant Breeding: Principles and Methods. Kalyani Publishers, India.

<b>Course Title</b>	:	Introduction to Animal Breeding
Course No.	:	AAP 121
<b>Credit Hours</b>	:	3 (2-0-1)

#### UNIT I

Population and Population Genetics; Gene and Genotypic frequency. Hardy- Weinberg Law; Hardy Weinberg Equilibrium; Proof of Hardy Weinberg Equilibrium; Stochastic and Deterministic Forces acting on Population; Mutation; Migration; Selection.

#### <u>UNIT II</u>

Dissection of Phenotype into its components;s Breeding Value: Definition, concept; Heritability: Definition, Concept, Estimation of heritability from regression of offspring to parents; Repeatability: Definition, Concept and estimation; Correlated traits: Phenotypic and Genetic correlation, Environmental correlation; Selection: Methods of Selection; Basis of selection; Selection Differential; Response to Selection; Correlated Response to Selection.

#### <u>UNIT III</u>

Animal Breeding: Aims of Animal Breeder; Natural vs Artificial Insemination; Mating Systems: Inbreeding, Out Breeding and Random mating; Breeding strategies in large ruminants (cattle, buffalo), small ruminants (sheep, goat) and swine; Poultry breeding; Lab animal breeding; Breed improvement programs conducted in India.

#### Practical

Estimation of Gene and Genotypic frequency; Chi-squared test for determining goodness of fit for HW-equilibrium; Estimation of heritability: regression of offspring on parents; Estimation of repeatability; Phenotypic correlation, genetic correlation, environmental correlation; Estimation of Selection Differential, Response to selection, Selection index. Linkage analysis from pedigree data.

#### **Suggested Readings**

Brah GS. 2014. Animal Genetics: Concepts and Implications. 2<sup>nd</sup> Ed. Kalyani Publishers. Nicholas FW. 2010. Introduction to Veterinary Genetics. 3<sup>rd</sup> Ed. Wiley-Blackwell Publication.

Richard M. Bourdon. 1999. Understanding Animal Breeding. 2<sup>nd</sup> Ed. Prentice Hall.

<b>Course Title</b>	:	Plant Tissue Culture
Course No.	:	BTO 121
<b>Credit Hours</b>	:	3 (2-0-1)

#### <u>UNIT I</u>

History of plant tissue culture; concept of totipotency; Concept of aseptic culture practices; Components of *in vitro* culture media and role of different macro and micro nutrients, vitamins, plant growth regulators and growth supplements; Sterilization techniques.

#### <u>UNIT II</u>

Various plant cell, tissue and organ culture techniques and uses; Somatic cell cultures; morphogenesis: organogenesis and somatic embryogenesis; Micropropagation: *In vitro* grafting, meristem culture; Anther, pollen, embryo, ovule, ovary culture; Protoplast culture and somatic hybridization; Somaclonal variation.

#### Practical

Good laboratory practices; Media preparation and sterilization; Surface sterilization of explants; Establishment of callus/cell suspension cultures; Micropropagation; Embryo culture; Anther and pollen culture; Induction of plant regeneration; Hardening and transfer to soil.

#### **Suggested Readings**

Bhojwani SS & Razdan MK. 1996. Plant Tissue Culture: Theory and Practice. Elsevier.
Bhojwani SS & Dantu PK. 2013. Plant Tissue Culture: An Introductory Text. Springer
Dixon RA & Gonzales RA. 2003. Plant Cell Culture: A Practical Approach. Oxford University press.

Helgason CD & Miller CL. 2005. Basic Cell Culture Protocols. 3rd Ed. Humana Press.

<b>Course Title</b>	:	<b>Molecular Biology</b>
Course No.	:	BTM 121
<b>Credit Hours</b>	:	3 (2-0-1)

#### <u>UNIT I</u>

History of molecular biology; Central dogma of life; Structure of DNA and RNA; Gene structure and function; DNA replication; transcription; Genetic code and translation; Structure of prokaryotic and eukaryotic nuclear and organelle genomes; Gene regulation in prokaryotes: Lac operon concept, tryp concept.

#### <u>UNIT II</u>

Introduction to microbial genetics; conjugation, transformation and transduction; Tools in molecular biology: Role of enzymes in molecular biology; Principles of Polymerase Chain Reaction; Electrophoresis; PCR and hybridization based molecular markers. \

#### Practical

Study of DNA structure (Experiments of Griffith, Avery et al, Hershey & Chase, Watson and Crick; Chargaff's rule). Study of chromosome structure. DNA Replication, repair and recombination. Study of mutations. Gene Mapping. Isolation and purification of plant and animal DNA; Measurement of nucleic acid concentration using spectrophotometer and gel electrophoresis; Preparation of bacterial competent cells, transformation and size estimation of plasmid DNA.

#### **Suggested Reading**

Allison LA. 2011. Fundamental Molecular Biology. Wiley Global Education.

- Carson S, Miller HB & Witherow DS. 2012. *Molecular Biology Techniques A Classroom Laboratory manual*. Elsevier.
- Kreuzer H & Massey A. 2008. Molecular Biology and Biotechnology: A Guide for Teachers. ASM Press.
- Lodish H, Berk A, Kaiser CA, Krieger M, Bretscher A, Ploegh H, Amon A & Scott MP. 2012. *Molecular Cell Biology*. W. H. Freeman.
- Sambrook J, Russel D. 2001. *Molecular Cloning: A Laboratory Manual*. 3rd Ed Cold Spring Harbor Laboratory Press.
- Surzycki S. 2000. Basic Techniques in Molecular Biology. Springer Berlin Heidelberg
- Voet D, Voet JG & Pratt CM. 2004. *Fundamentals of Biochemistry*. 2nd Ed. New York: Wiley.
- Walker JM & Rapley R. 2000. *Molecular Biology and Biotechnology*. 4th Ed. The Royal Society of Chemistry.
- Watson JD, Bakee TA, Bell SP, Gann A, Levine M & Losick R. 2008. *Molecular Biology of the Gene*. 6th Ed. Pearson Education International.

<b>Course Title</b>	:	<b>Biodiversity and its Conservation</b>
Course No.	:	BTC 121
<b>Credit Hours</b>	:	2 (2-0-0)

#### <u>UNIT I</u>

Concepts of biodiversity, bioresource and wildlife management, conservation strategies: *in situ* and *ex situ* conservation; Wild life conservation projects in India; Protection of biodiversity for its suitable utilization; Threats to biodiversity; WCU Red data book; Biodiversity hotspots in India; National bureaus of genetic resources.

#### <u>UNIT II</u>

Sustainable development; Diversification of cropping system; Diversity of indigenous livestock; Vulnerability and extinction of flora and fauna; Endangered species in various ecosystems; Germplasm banks; Environmental impact assessment; Bioremediation and biosafety; Introduction to regulatory agencies and legislation.

#### **Suggested Readings**

- Das MK & Choudhury BP. 2008. A Text book on Plant Nomenclature and Biodiversity Conservation. Kalyani Publishers.
- Hopsetti BB. & Venketashwarlaru M. 2001. Trends in Wild Life Conservation and Management. Vol. 2, Daya Publishing House.

Singh MP & Singh BS. 2002. Plant Biodiversity and Taxonomy. Daya Publishing House, Delhi.

<b>Course Title</b>	:	Microbiology
Course No.	:	BTP 121
<b>Credit Hours</b>	:	3 (2-0-1)

#### UNIT I

History of Microbiology-its applied areas; Microorganisms and their role in fermentation; Germ theory of diseases and protection; Introduction to eukaryotic and prokaryotic cell; Major groups of eukaryotes- fungi, algae and protozoa; Major groups of prokaryotes – Actinomycetes, Cyanobacteria, Archaebacteria, Rickettsias and Chlamydia; Preservation of microorganisms; Microbial repositories at national and international level.

#### <u>UNIT II</u>

Bacterial growth; Metabolism in bacteria- ATP generation, chemoautotrophy, photoautotrophy, respiration, fermentation; Viruses: Bacteriophages - structure and properties, lytic and lysogenic cycles; viriods, prions.

#### <u>UNIT III</u>

Microbial groups in soil; Microbes in biotic and abiotic stressed environments; Microbial transformation of carbon, nitrogen and sulphur; Biological nitrogen fixation; Beneficial microorganisms in agriculture-biofertilizers, microbial pesticides; Plant microbe interaction; Microbes in composting and biodegradation; Microbiology of water and food.

#### Practical

Microscope and other instruments in a microbiological laboratory; Media preparation, sterilization and aseptic methods for isolation, identification, preservation and storage; Identification of bacteria by staining methods; Enumeration of bacteria by pour plate and spread plate methods; Micrometery.

#### **Suggested Readings**

Brock TD. 1961. Milestones in Microbiology. Infinity Books.

Pelczar MJ, Chan ECS & Kreig NR. 1997. *Microbiology: Concepts and Application*. Tata McGraw Hill.

Stainier RY, Ingraham JL, Wheelis ML & Painter PR. 2003. General Microbiology. MacMillan.

Tauro P, Kapoor KK & Yadav KS. 1996. Introduction to Microbiology. Wiley Eastern.

#### SEMESTER-III

<b>Course Title</b>	:	Biomathematics
Course No.	:	BAS 211
<b>Credit Hours</b>	:	3 (2-0-1)

#### <u>UNIT-I</u>

Rolle's theorem; Lagrange's theorem; Taylor's and Maclaurin's series; Partial differentiation, Euler's theorem on homogeneous function, change of variable; Jacobian, maxima and minima of two or more than two variables eigen values and eigen vectors of a matrix; Reduction formulae, definite integrals and its applications.

#### <u>UNIT-II</u>

Solution of ordinary differential equation of first degree and first order and their application for determination of volume of blood and drug distribution; Epidemic models, Simultaneous differential equation of first order and their applications to predator models; Linear differential equations of higher order and their applications to simple biological problem; Numerical methods for solving algebraic and transcendental equations.

#### Practical

Tutorials on Taylor's and Maclaurin's expansions; Partial differentiation; Euler's theorem; Change of variable, total derivative, implicit function, maxima and minima, eigen values and eigen vectors of matrix, reduction formulae, definite integrals and their properties; Epidemic models, predator models; Determination of volume of blood and drug distribution; Ordinary differential equation of first order, linear differential equation of higher older and their applications to biological problems, numerical methods.

#### **Suggested Readings**

Grewal BS. 2013. *Higher Engineering Mathematics*. Khanna Publishers, India. Rastogi SK. 2008. *Biomathematics*. Krishna Prakashan Media Pvt. Ltd.

Srivastava AC & Srivastava PK. 2011. *Engineering Mathematics*. Vol.I. PHI Learning Pvt. Ltd.

Srivastava AC & Srivastava PK. 2011. *Engineering Mathematics*. Vol.III . PHI Learning Pvt. Ltd.

<b>Course Title</b>	:	Information and Communication Technology
Course No.	:	BAS 214
<b>Credit Hours</b>	:	2 (1-0-1)

#### <u>UNIT I</u>

IT and its importance; IT tools; IT-enabled services and their impact on society; Computer fundamentals; Hardware and software; Input and output devices; Word and character representation.

#### <u>UNIT II</u>

Features of machine language, assembly language, high-level language and their advantages and disadvantages; Principles of programming - algorithms and flowcharts.

#### <u>UNIT III</u>

Operating systems (OS) - definition, basic concepts; Introduction to WINDOWS and LINUX Operating Systems; Local area network (LAN); Wide area network (WAN); Internet and World Wide Web; HTML and IP.

#### UNIT IV

Introduction to MS Office - Word, Excel, Power Point; Audio visual aids - definition, advantages, classification and choice of A.V. aids; Criteria for selection and evaluation of A.V aids; Video conferencing; Communication process, Berlo's model, feedback and barriers to communication.

#### Practical

Exercises on binary number system; Algorithm and flow chart; MS Word; MS Excel; MS Power Point; Internet applications: web browsing, creation and operation of email account; Analysis of data using MS Excel; Handling of audio visual equipments; Planning, preparation, presentation of posters, charts, overhead transparencies and slides; Organization of an audio visual programme.

#### **Suggested Readings**

Gurvinder Singh, Rachhpal Singh & Saluja KK. 2003. Fundamentals of Computer Programming and Information Technology. Kalyani Publishers.

- Harshawardhan P. Bal. 2003. Perl Programming for Bioinformatics. Tata McGraw-Hill Education.
- Kumar A 2015. Computer Basics with Office Automation. IK International Publishing House Pvt Ltd.

Rajaraman V & Adabala N. 2015. *Fundamentals of Computers*. PHI Recommended Latest Online Tutorials (over Internet).

<b>Course Title</b>	:	Economics and Marketing
Course No.	:	AAE 212
<b>Credit Hours</b>	:	3 (2-0-1)

#### <u>UNIT I</u>

Economics – Terms and definitions; Consumption, demand, price and supply; Factors of production; Gross Domestic Product; Role of Biotechnology/ Agriculture sector in national GDP.

#### <u>UNIT II</u>

Marketing – definition; Marketing process; Need for marketing; Role of marketing; Marketing functions; Classification of markets; Marketing of various channels; Price spread; Marketing efficiency; Constraints in marketing of agricultural produce; Market intelligence.

#### <u>UNIT III</u>

Basic guidelines for preparation of project reports; Bank norms; Insurance; SWOT analysis; Crisis management.

#### Practical

Techno-economic parameters for preparation of projects; Preparation of bankable projects for various biotechnology/ agricultural products and value added products; Identification of marketing channel; Calculation of price spread; Identification of market structure; Visit to different markets, market institutions; Study of SWC, CWC and STC; Analysis of information of daily prices; Marketed and marketable surplus of different commodities.

#### **Suggested Readings**

Acharya SS & Aggarwal NL. 2011. *Agricultural Marketing in India*. Fifth Edition. Oxford and IBH Publishing Company Pvt. Ltd.

Ahuja HL. 2007. *Advanced Economic Theory*. S Chand and Company. Chandra P. 1984. *Projects: Preparation, Appraisal & Implementation*. McGraw Hill Inc. Dewett KK. 2005. *Modern Economic Theory*. S Chand and Company. Gupta RD & Lekhi RK. 1982. *Elementary Economic Theory*. Kalyani Publishers. Sampat Mukherjee. 2002. *Modern Economic Theory*. New Age International.

<b>Course Title</b>	:	Fundamentals of Crop Protection
Course No.	:	ENT/PPA 212
<b>Credit Hours</b>	:	3 (2-0-1)

#### <u>UNIT I</u>

Insects - their general body structure; Importance of insects in agriculture; Life cycle of insects; Insects diversity; Feeding stages of insects and kinds (modifications) of mouth parts; Concepts in population build-up of insects – GEP, DB, EIL, ETH and pest status; Causes of insect-pests out break; General symptoms of insects attack; Principles and methods of insect-pests management; Integrated Pest Management concept; Bioecology and management of important pests of major crops and storage products.

#### <u>UNIT II</u>

Importance and scope of plant pathology; Concept of disease in plants; Nature and classification of plant diseases; Importance and general characters of fungi, bacteria, fastidious bacteria, nematodes, phytoplasmas, spiroplasmas, viruses, viroids, algae, protozoa and phanerogamic parasites; Pathogenesis due to obligate and facultative parasites; Variability in plant pathogens; Conditions necessary for development of disease epidemics; Survival and dispersal of plant pathogens; Management of key diseases and nematodes of major crops.

#### Practical

Familiarization with generalized insect's body structure and appendages; Life stages; Acquaintance with insect diversity; Identification of important insect-pests of cereals, cotton, oilseeds, pulses, sugarcane, fruit and vegetables crops and stored-grains, and their symptoms of damage; Acquaintance with useful insects: predators, parasitoids, pollinators, honey bees and silk worms; Acquaintance with various pesticidal formulations; Principles and working of common plant protection appliances; Calculation for preparing spray material; Acquaintance to plant pathology laboratory equipment; of culture media for fungi and bacteria; Demonstration of Koch's Preparation postulates; Study of different groups of fungicides and antibiotics and methods of their evaluation; Diagnosis and identification of important diseases of cereals, cotton, oilseeds, pulses, sugarcane, fruit and vegetables crops and their characteristic symptoms.

#### **Suggested Readings**

Agrios, GN. 2010. Plant Pathology. Acad. Press.

Atwal AS & Dhaliwal GS. 2002. Agricultural Pests of South-Asia and Their Management. Kalyani Publishers.

Dhaliwal GS & Arora R. 1996. Principles of Insect Pest Management. National Agriculture Technology Information Centre.

Dhaliwal GS, Singh R & Chhillar BS. 2006. Essentials of Agricultural Entomology. Kalyani Publishers.

- Mehrotra RS & Aggarwal A. 2007. Plant Pathology. 7th Ed. Tata Mc Graw Hill Publ. Co. Ltd.
- Singh H. 1984. House-hold and Kitchen Garden Pests Principles and Practices. Kalyani Publishers.

Singh RS. 2008. Plant Diseases. 8th Ed. Oxford & IBH. Pub. Co.

Singh RS. 2013. Introduction to Principles of Plant Pathology. Oxford and IBH Pub. Co. Stakman EC & Harrar JG. 1957. Principles of Plant Pathology. Ronald Press, USA.

Tarr SAJ. 1964. The Principles of Plant Pathology. McMillan, London.

Vander Plank, JE. 1975. Principles of Plant Infection. Acad. Press.

Course Title	:	Livestock Product Technology
Course No.	:	LPT 211
<b>Credit Hours</b>	:	3 (2-0-1)

#### UNIT I

Composition and nutritive value of milk and factors effecting composition of milk; Physiochemical properties of milk; Determination of microbial load in milk and milk products; Milk Processing: Collection, chilling, standardization, pasteurization and homogenization; Toxins and pesticide residues in milk and milk products; Organic milk food products; Bureau of Indian Standards for milk and milk products; Sanitation in milk plant.

#### UNIT II

Retrospect and prospects of meat industry in India; Structure and composition of muscle (including poultry), nutritive value of meat, Meat adulteration, preservation of meat, Physico – chemical and microbiological quality of meat and meat products. Laws governing national, international trade in meat and meat products, organic meat food products, food products of genetically modified animals.

#### Practical

Sampling of milk, estimation of fat, solids not fat (SNF) and total solids, Platform tests, cream separation, Microbiological quality of milk, meat and meat products. Chilling/freezing of meat, meat products, preservation of meat and meat products. Visit to modern milk and meat processing units.

#### **Suggested Readings**

Aberle ED, Forrest JC, Gerrard DE & Mills EW. 2012. Principles of Meat Science.5<sup>th</sup> Eds. Kendall Hunt Publishing

Ledward DA & Lawrie RA. 2006. Lawrie's Meat Science, 7th Eds. Woodhead Publishing

Sharma BD.1999. Meat and Meat Products Technology: Including Poultry Products Technology. Jaypee Bros. Medical Publishers

Sukumar De. 2001. Outlines of Dairy Technology. Oxford University Press.

Varnam A& Sutherland JP. 2001. *Milk and Milk Products: Technology, Chemistry and Microbiology*. Springer Science & Business Media

<b>Course Title</b>	:	<b>Breeding of Field Crops</b>
Course No.	:	GPB 212
<b>Credit Hours</b>	:	3 (2-0-1)

#### <u>Unit I</u>

Application of genetic, cytogenetic and biotechnological techniques in breeding of: Wheat, triticale, rice, maize, bajra, barley, sorghum, cotton, sugarcane, important pulses, oilseeds and forage crops including their origin and germplasm sources.

#### <u>Unit II</u>

Problems and present status of crop improvement in India with emphasis on the work done in state National and International centres of crop improvement.

#### <u>Unit III</u>

Classes of seed; seed production and maintenance; seed storage; seed certification.

#### Practical

Emasculation and hybridization techniques; Handling of segregating generations : pedigree method, bulk method, back cross methods; Field layout of experiments; Field trials, maintenance of records and registers; Estimation of heterosis and inbreeding depression; Estimation of heritability; Parentage of released varieties/hybrids; Study of quality characters; Sources of donors for different characters; seed sampling; seed quality; seed viability; seed vigour; seed health testing; Visit to seed production plots.

#### Suggested Readings

Chopra VL 2001. Breeding Field Crops. Oxford and IBH Publishing Co.

Fehr WR. 1987. Principles of Cultivar Development, Vol. II Crop Species. MacMillan Publishing Co.

Sleper DA & Poehlman JM. 2006. Breeding Field Crops. Wiley-Blackwell.

<b>Course Title</b>	:	Animal Health Care
Course No.	:	VMD/VMC 212
<b>Credit Hours</b>	:	3 (2-0-1)

#### <u>UNIT I</u>

Introduction to animal health; Animal Vaccinations; history of disease diagnoses and medicine; classification of diseases; Introduction to fore stomach disorders in ruminants.

#### <u>UNIT II</u>

Introduction to important diseases of respiratory, urinary, musculoskeletal and cardiovascular system of domestic animals. Introduction to common metabolic, bacterial, viral, parasitic and blood protozoan diseases of domestic animals. Importance of animal health in relation to public health.

#### Practical

Introduction to veterinary hospital; methods of sample collection; introduction to common disease diagnostic tests in animals; vaccination schedule in domestic animals. Microscopic examination of parasites. Clinical diagnostics: urine, blood, milk, sputum, faeces examination.

#### **Suggested Readings**

Blood DC & Henderson JA. 1968. Veterinary Medicine. Bailliere Tindall publishers.

- Bradford P & Smith DVM. 2014. Large Animal Internal Medicine. 5<sup>th</sup> edition. Mosby Publishers.
- Chakrabarti Amalendu. 2007. Textbook of Clinical Veterinary Medicine. Kalyani Publishers.
- Stephen J, Ettinger DVM, Edward C & Feldman DVM. 2010. *Textbook of Veterinary Internal Medicine Expert Consult.* 7<sup>th</sup> edition. Saunders Publishers.

<b>Course Title</b>	:	Livestock Production and Management
Course No.	:	LPM 212
<b>Credit Hours</b>	:	3 (2-0-1)

#### UNIT I

Livestock history in India: Vedic, medieval and modern era; Demographic distribution of livestock and role in economy; Introductory animal husbandry; Breeds of livestock; Cattle, Buffalo, Sheep, Goat and Pig; Important traits of livestock; General management and feeding practices of animals; Handling and restraining of animals; Housing systems. Importance of grasslands and fodders in livestock production; Common farm management practices including disinfection, isolation, quarantine and disposal of carcass; Common vices of animals and their prevention; Diseases and parasite control & hygiene care.

# <u>UNIT II</u>

History and economic importance of poultry; Poultry breeds; Reproductive system of male and female birds; Formation and structure of eggs; Important economic traits of poultry, Egg production, Egg weight, Egg quality; Fertility and Hatchability, Plumage characteristics and comb types.

Care and management of chicks, grower and layers/broiler; Brooding management; Hatchery practices; Poultry Diseases, control and hygiene care;

# Practical

Visit to livestock farms/demonstration centres; Breeds of cattle, buffalo, sheep, goat and Pigs; Familiarization with body parts of animals; Handling and restraining of cattle, buffalo, sheep, goat and swine; Male and female reproductive system and Artificial Insemination; Feeding of livestock; Methods of identification: marking, tattooing, branding, tagging; Milking methods; Record Keeping.

Visit to the Poultry farm; Poultry breeds; Body parts of chicken, duck, quail and turkey; Housing, equipment, nesting and brooding requirements; Male and female reproductive system; Methods of identification and sexing; Hatchery layout and equipment; Identification of diseases and control of parasites, Vaccination; Maintenance of farm records;

# **Suggested Readings**

Banerjee GC. 1989. Text Book of Animal Husbandry. Oxford and IBH.
ICAR. 1962. Handbook of Animal Husbandry. ICAR Publication.
Parsad Jagdish. 2001. Poultry Production and Management. Kalyani Publishers.
Sastry NSR & Thomas CK. 1991. Dairy Bovine Production. Kalyani Publishers.
Singh RA. 1990. Poultry Production. Kalyani Publishers.
Thomas CK & Sastry NSR. 2013. Livestock Production Management. Kalyani

Publishers.

<b>Course Title</b>	:	<b>Recombinant DNA Technology</b>
Course No.	:	BTR 211
<b>Credit Hours</b>	:	3 (2-0-1)

# <u>UNIT I</u>

Recombinant DNA technology; Restriction endonucleases: Types and uses; DNA ligases; Vectors: plasmids, cosmids, phagemids, BACs, PACs, YACs, transposon vectors, expression vectors, shuttle vectors, binary plant vectors, co-integrating vectors.

#### <u>UNIT II</u>

Competent cells; Gene isolation and cloning; Genetic transformation of *E. coli*; Gel electrophoresis; Preparation of probes; Southern blotting; Northern blotting; Western blotting; PCR and gene amplification.

## Practical

Orientation to recombinant DNA lab; preparation of stock solutions and buffers; Plasmid DNA isolation; Genomic DNA isolation; Quality and quantity determination of DNA;

restriction digestion of DNA; Agarose gel electrophoresis, SDS-PAGE; PCR; Genetic transformation of *E. coli*; Screening of recombinant DNA clones in *E. coli*.

# **Suggested Readings**

Brown TA. 1998. Genetics: A Molecular Approach. 3rd Ed. Stanley Thornes.
Singer M & Berg P. 1991. Genes & Genome. University Science Books.
Winnacker EL. 2003. From Genes to Clones: Introduction to Gene Technology. 4th Ed. Panima Publishers.

Watson JD & Zoller M. Recombinant DNA. 3rd Ed. Panima Publishers

<b>Course Title</b>	:	Plant Physiology
Course No.	:	<b>BTB 211</b>
<b>Credit Hours</b>	:	3 (2-0-1)

# <u>UNIT I</u>

Plant physiology, its scope in agriculture; Osmosis, imbibition, water absorption, water translocation and transpiration; Stomatal mechanisms; Physiological role and deficiency symptoms of major and minor elements, Absorption and translocation of minerals.

#### UNIT II

Concepts of photosynthesis, photorespiration, respiration and translocation of photoassimilates; Dynamics of growth; Stress physiology; Nitrogen and sulphur metabolism; Plant growth regulators: Their biosynthesis and physiological roles, seed germination & seed dormancy, senescence, vernalization.

#### Practical

Demonstration of processes of diffusion, osmosis, imbibition and plasmolysis; Ascent of sap, transpiration; Deficiency symptoms of nutrients in crop plants; Plant growth analysis; Quantitative and qualitative estimation of plant pigments; Experiments on photosynthesis and respiration; Effects of plant growth regulators on plant growth and seed germination; Experiments on seed dormancy; Relative water content and plant water potential; Proline estimation.

# **Suggested Readings**

Bhatia KN & Prashar AN. 1990. *Plant Physiology*. Trueman Book Company. Salisbury FB. & Ross CW. 1992. *Plant Physiology*. Wordsworth Publishing Company. Srivastava HN. 2000. *Plant Physiology*. Pradeep Publications.

#### **SEMESTER-IV**

<b>Course Title</b>	:	Biophysics
Course No.	:	<b>BAS 221</b>
<b>Credit Hours</b>	:	3 (2-0-1)

# <u>UNIT I</u>

Quantum mechanics; Electronic structure of atoms; The wave particle duality, wave length of de-Broglie waves; Phase and group velocity; Some basic concepts of quantum mechanics; Schrodinger's wave equations; Particle in a box; Quantum mechanical tunneling; Ist and IInd law of thermodynamics; Enthalpy; Entropy; Statistical and thermodynamic definition of entropy; Helmholtz free energy, Equilibrium thermodynamic; Near-equilibrium thermodynamic; Gibbs free energy; Chemical potential; Thermodynamic analysis of membrane transport.

# <u>UNIT II</u>

Hydration of macromolecules; Role of friction; Diffusion; Sedimentation; The ultracentrifuge; Viscosity; Rotational diffusion; Light scattering, Small angle x-ray scattering; Ultraviolet and visible spectroscopy; Circular dichroism(CD) and optical rotatory dispersion(ORD); Fluorescence spectroscopy; Infrared spectroscopy; Raman spectroscopy; Electron spin resonance; NMR spectroscopy; Light microscopy.

#### <u>UNIT III</u>

Electron optics; Transmission electron microscope (TEM); Scanning electron microscope(SEM); Preparation of the specimen for electron microscopy; Image reconstruction; Electron diffraction; Tunnelling electron microscope; Atomic force microscope; Crystals and symmetries, crystal systems, point group and space groups; Growth of crystals of biological molecules; X-ray diffraction.

## Practical

Refractive index and dispersive power of the prism using spectrometer; Calibration of prism spectrometer; Newton's rings; Polarimeter; Diffraction grating; Resolving power of telescope and grating; Ostwald viscometer; Planck's constant using photovoltaic cell; Photospectrometer; Photoelectric effect; Stefan's constant; Thermal diffusivity in metals.

# Suggested Readings

Chang R. 2005. *Physical Chemistry for the Biosciences*. University Science Books.
Glaser. 2012. *Biophysics*. Springer.
Pattabhi V & Gautam N. 2002. *Biophysics*. Narosa Publishing House.
Rodney Cotterill. 2002. *Biophysics: An Introduction*. John Wiley & Sons.
Srivastava PK. 2006. *Elementary Biophysics: An Introduction*. Narosa Publishing House.

<b>Course Title</b>	:Entre	epreneurship Development and Business Management
Course No.	:	AEC/AAE 222
<b>Credit Hours</b>	:	2 (1-0-1)

#### <u>UNIT I</u>

Concept of entrepreneur; Entrepreneurship development; Assessment of entrepreneurship skills; SWOT analysis and achievement motivation; Entrepreneurial behaviour; Government policy and plan for entrepreneurship development; Setting up of a new entrepreneurial venture; Environmental factors influencing entrepreneurship; Constraints in setting up of agro based industries;

#### UNIT II

Definition of business; Value chain concept in business; Stakeholders in business; Stages of Indian business; Importance of agribusiness in Indian economy and factors ransforming Indian agribusiness; Government as a regulatory body in agribusiness; Opportunities and challenges to Indian agribusiness.

## <u>UNIT III</u>

Management: Definition, importance and functions; Levels of management; Planning: Definition, steps in planning, types of plan; Organizing: Meaning of organizing and organization; Developing leadership skills; Encoding and decoding communication skills; Developing organizational and managerial skill; Problem solving skill; Supply chain management and total quality management; Project planning, formulation and report preparation.

# Practical

Preparation of project report for starting a new venture; Case studies of successful entrepreneurs, analysis and discussion; Preparation of complete marketing plan of selected product/service; Case studies related to project management; Visits to industrial and agri-business houses; Numerical problems; Preparation of project report for various business ventures.

## **Suggested Readings**

Harold Koontz & Heinz Weihrich. 2004. *Essentials of Management: An International Perspective*, 2nd Ed. Tata Mc-Graw Hill Publishing Pvt Ltd.

Mukesh Pandey & Deepali Tewari. 2010. The Agribusiness Book. IBDC Publishers.

Nandan H. 2011. Fundamentals of Entrepreneurship. PHI Learning Pvt Ltd India.

Philip Kotler, Kavin Lane Keller, Abraham Koshy & Mithileshwar Jha. 2012. *Marketing Management: A South Asian Perspective*. Pearson Education.

Poornima Charantimath. 2006. Entrepreneurship Development: Small Business Enterprise. Pearson Education.

Stephans P Robbins & Mary Coulter. 2003. Management. Pearson Education.

<b>Course Title</b>	:	Electronics and Instrumentation in Biotechnology
Course No.	:	BTB/AGE 221
<b>Credit Hours</b>	:	2 (1-0-1)

# <u>UNIT I</u>

Electronics; PN junction diode, diode forward and reverse characteristics; Diode as a circuit element; Application of PN junction diode such as: half wave, full wave bridge rectifier, clipper, clamper and voltage multiplier circuit; Construction and working of bipolar transistor, load line concept, analysis and design of various biasing methods of NPN transistor with common emitter configuration; AC model and analysis of small signal NPN transistor with common emitter configuration; Concept of generalized instrumentation system; Transducers for the measurement of temperature using thermometer and thermocouple, linear displacement measurement using LVDT; Force measurement using the strain gauge.

# <u>UNIT II</u>

Principles and working of laboratory equipments: Table top, refrigerated and ultra centrifuges; Laminar air flow; Autoclaves, pH meter; Fermenters; Temperature control shakers, BOD shakers; Gel electrophoresis, 2-D gel electrophoresis, gel documentation, gel driers; ELISA readers; Freeze driers/lypholizers; Spectrophotometers; Gene pulser;

Particle gun; Plant growth chambers; Thermal cyclers; Realtime PCR; DNA synthesizer; DNA sequencer; Microscopes: Light, stereo, phase contrast and inverted.

# Practical

To familiarize laboratory equipment and its equipment working; Forward and reverse VI Characteristics of a PN junction diode; To study half wave, full wave and bride rectifier using diode; Clipper, Clamper and Voltage multiplier circuit; To determine input V-I Characteristics s of bipolar transistor for common emitter configuration; To determine output V-I Characteristics s of bipolar transistor; To design and test a biasing circuits for CE transistor; To design and test a biasing circuits for CE transistor; To study the measure of temperature using the available sensor; To measure displacement with the available sensor; To study force with the available sensor.

# **Suggested Readings**

Edward William Golding & Frederick Charles Widdis. 1969. Electrical Measurements and Measuring Instruments. Pitman.
Gupta JB.2009. Basic Electronics. S. K. Kataria & Sons.
Malvino. 2007. Electronics Principles. Tata McGraw-Hill Education.
Manhas P. & Thakral S. 2010. Digital Electronics. S. K. Kataria & Sons.
Sharma Sanjay. 2012. Electronics Devices & Circuits. S. K. Kataria & Sons.

<b>Course Title</b>	:	<b>General Biochemistry</b>
Course No.	:	BTB 222
<b>Credit Hours</b>	:	4 (3-0-1)

# <u>UNIT I</u>

Introduction and importance; Cell structure; Bio molecules: Carbohydrates, lipids, proteins and nucleic acids - structure, functions and properties; Enzymes: Classification, factors affecting activity; Structure and role of water in biological system; Acids, bases and buffers of living systems; The pK of biomolecules; Vitamins and hormones.

# <u>UNIT II</u>

Bioenergetics; Metabolism - basic concept: Glycolysis, Citric acid cycle, Pentose phosphate pathway, Oxidative phosphorylation, Fatty acid oxidation; General reactions of amino acid degradation; Biosynthesis - carbohydrates, lipids, proteins, nucleic acids.

# <u>UNIT III</u>

Secondary metabolites: Terpenoids, alkaloids, phenolics and their applications in food and pharmaceutical industries.

# Practical

Qualitative tests for carbohydrates, amino acids, proteins and lipids; Extraction and characterization of lipids by TLC; Determination of acid, iodine and saponification values of oil; Extraction, quantitative estimation and separation of sugars by paper chromatography; Determination of phenols; Determination of free amino acids and proteins.

# Suggested Readings

Berg JM, Tymoczko JL, & Stryer L. 2002. Biochemistry. 5th Ed. W.H. Freeman & Co.

Com EE & Stumpf PK. 2010. Outlines of Biochemistry. 5th Ed. John Wiley Publications.

Goodwin, TW & Mercer EI. 1983. Introduction to Plant Biochemistry. 2nd Ed. Oxford, New York. Pergaman Press.

Murray RK, David B., Botham KM & Kennelly PJ. 2012. Harper's Illustrated Biochemistry. 29th Ed. Lange Medical Books/Mc. Graw Hill.

- Nelson DL & Cox MM. 2000. Lehninger Principles of Biochemistry. 5th Ed. C.B.S Publilshers, Prentice Hall.
- Wilson K & Walker J. 1994. Principles and Techniques of Biochemistry and Molecular Biology. 7th Ed. Cambridge University Press.

<b>Course Title</b>	:	Introductory Bioinformatics
Course No.	:	BTI/BTR 221
<b>Credit Hours</b>	:	3 (2-0-1)

# <u>UNIT I</u>

Introduction to bioinformatics; Development and scope of bioinformatics; Applications of computers in bioinformatics: Operating systems, hardware, software, Internet, www resources, FTP.

# <u>UNIT II</u>

Primary databases: Nucleotide sequence databases (GenBank, EMBL), protein sequence databases; Secondary databases: SwissProt/TrEMBL, conserved domain database, Pfam;

Structure databases: Protein Data Bank (PDB), MMDB, SCOP, CATH; File formats: Genbank, EMBL, Fasta, PDB, Flat file, ASN.1, XML.

# <u>UNIT III</u>

Introduction to sequence alignment and its applications: Pair wise and multiple sequence alignment, concept of local and global alignment; Algorithms: Dot Matrix method, dynamic programming methods (Needleman–Wunsch and Smith–Waterman); Tools of MSA: ClustalW, TCoffee; Phylogeny; Introduction to BLAST and FASTA.

# Practical

Basic computing: Introduction to UNIX, LINUX; Nucleotide information resource: EMBL, GenBank, DDBJ, Unigene; Protein information resource: SwissProt, TrEMBL, Uniprot; Structure databases: PDB, MMDB; Search Engines: Entrez, ARSA, SRS; Similarity Searching: BLAST and interpreting results; Multiple sequence alignment: ClustalW; Structure visualization of DNA and proteins using Rasmol.

# **Suggested Readings**

Baxevanis AD. & Ouellette BFF. 2001. Bioinformatics: A practical guide to the analysis of genes and proteins. John Wiley and Sons.

Mount DW. 2001. Bioinformatics: Sequence and Genome Analysis. Cold Spring Harbor. Xiong J.2006. Essential Bioinformatics. Cambridge University Press.

<b>Course Title</b>	:	Plant Genetic Transformation
Course No.	:	BTM 221
<b>Credit Hours</b>	:	3 (2-0-1)

# <u>UNIT I</u>

History of plant genetic transformation; Generation of gene construct and maintenance; Genetic transformation: *Agrobacterium* mediated, biolistics, electroporation, liposome, Polyethylene glycol, *in planta* methods.

#### UNIT II

Selection and characterization of transgenic plants using selectable and reportable markers; PCR; qRT-PCR; Southern, Northern, ELISA and Western techniques; Application of genetic transformation: for quality, yield, biotic, and abiotic stresses; Biosafety aspects of transgenic plants and regulatory framework.

## Practical

Preparation of stock solutions, Preparation of competent cells of Agrobacterium tumefaciens; Restriction mapping of plasmid, Construction of binary vector and its transfer to an Agrobacterium strain; Confirmation of transformed bacterial colonies; Agrobacterium tumefaciens mediated and biolistic plant transformation; Colony hybridization.

## Suggested Readings

Green & Sambrook. 2014. *Molecular Cloning: A Laboratory Manual*. 4<sup>th</sup> Ed. 3 Vol Sets. Cold Spring Harbor Laboratory Press.

Grierson D. 2012. Plant Genetic Engineering. Springer Netherlands.

Primose SB & Twyman RM. 2006. Principles of Gene Manipulation and Genomics, 7<sup>th</sup>Ed. Black Well Publishing.

Sambrook J, Russel D. 2001. *Molecular Cloning: A Laboratory Manual*. 3rd Ed Cold Spring Harbor Laboratory Press.

Stewart NC Jr. 2008. Plant Biotechnology and Genetics: Principles, Techniques and Applications. John Wiley & Sons Inc.

<b>Course Title</b>	:	<b>Classical and Molecular Cytogenetics</b>
Course No.	:	BTC 221
<b>Credit Hours</b>	:	3 (2-0-1)

#### UNIT I

Introduction and history; Mitosis and meiosis; Structure of chromatin; Chromosome structure and chromosome landmarks; Specialized chromosomes; Differential staining of the chromosomes- Q-banding, G banding, C banding, R banding; *In situ* hybridization-FISH, GISH.

#### UNIT II

Changes in chromosome number: aneuploidy- monosomy, trisomy and tetrasomy, haploidy and polyploidy- autopolyploidy and allopolyploidy; Methods of doubled

haploid production; Structural aberrations of chromosomes: deletions, duplications, inversions and translocations; Locating genes on chromosomes; Genome analysis.

# Practical

Preparation of chromosome stains; Pollen fertility; Preparation of mitotic and meiotic slides of plant/animal cells; Preparation of karyotypes; C/G banding of the chromosomes; Genomic *in situ* hybridization; Microphotography.

# **Suggested Readings**

Becker K & Hardin. 2004. *The World of Cell.* 5th Ed. Pearson Edu.
Carroll M. 1989. *Organelles*. The Guilford Press.
Charles B. 1993. *Discussions in Cytogenetics*. Prentice Hall.
Gupta PK. 2007. *Cytogenetics*. Rastogi publications.
Khush GS. 1973. *Cytogenetics of Aneuploids*. Academic Press.
Mahabal Ram. 2010. *Fundamentals of cytogenetics and genetics*. PHI Learning Pvt. Ltd.
Yao-Shan Fan. 2002. *Molecular Cytogenetics: Protocols and Applications*. Humana Press.

<b>Course Title</b>	:	<b>Microbial Genetics</b>
Course No.	:	BTP 221
<b>Credit Hours</b>	:	3 (2-0-1)

## <u>UNIT I</u>

Microorganisms as tools for genetic studies; Genetic variability in microorganisms; Genetic analysis of representative groups of bacteria, fungi and viruses; Random and tetrad spore analysis; Recombination and chromosomal mapping; Complementation - intergenic and intragenic.

#### <u>UNIT II</u>

Bacterial plasmids; Structure, life cycle, mode of infection and their role in genetic engineering; Transfer of genetic material in bacteria: Conjugation, transformation and transduction; Genetics of bacteriophage: T4, lambda and M13 - fine structure of gene, life cycle, mode of infection; Mutation: types, mutagens, DNA damage and repair; Transposable elements; Lac operon; Yeast genetics.

### <u>UNIT III</u>

Concept and application of recombinant DNA technology; Use of genetic tools to improve the microbial strains with respect to industry, agriculture and health.

#### Practical

Conjugation and transformation in bacteria; Spontaneous and auxotrophic mutation; Chemical and UV mutagenesis in fungi and bacteria; Complementation in fungi; Identification of mutants using replica plating technique; Isolation of genomic DNA from *E. coli*; Isolation and curing of plasmid;Identification of plasmid by electrophoresis / antibiotic plates.

# **Suggested Readings**

Birge EA. 1981. Bacterial and Bacteriophage Genetics. Springer Verlag.

Gardner JE, Simmons MJ & Snustad DP. 1991. Principles of Genetics. John Wiley& Sons.
Lewin B.1999. Gene. Vols. VI-IX. John Wiley & Sons.
Maloy A & Friedfelder D. 1994. Microbial Genetics. Narosa.
Scaife J, Leach D & Galizzi A 1985. Genetics of Bacteria. Academic Press.
William Hayes 1981. Genetics of Bacteria. Academic Press.

# SEMESTER-V

<b>Course Title</b>	:	<b>Agricultural Informatics</b>
Course No.	:	BAS 311
<b>Credit Hours</b>	:	3 (2-0-1)

#### <u>UNIT I</u>

Introduction to computers; Anatomy of computers; Memory concepts, units of memory; Operating system, definition and types; Applications of MS-Office for creating, editing and formatting a document; Data presentation, tabulation and graph creation; Statistical analysis, mathematical expressions; Database, concepts and types, creating database; Uses of DBMS in Agriculture; Internet and World Wide Web (WWW), concepts, components and creation of web; HTML & XML coding.

## <u>UNIT II</u>

Computer programming, concepts; Documentation and programme maintenance; Debugging programmes; Introduction to Visual Basic, Java, Fortran, C/ C++, etc.; Standard input/output operations; Variables and constants; Operators and expressions; Flow of control; Inbuilt and user defined functions; Programming techniques for agriculture.

# <u>UNIT III</u>

e-Agriculture, concepts, design and development; Application of innovative ways to use information and communication technologies (IT) in agriculture; ICT for data collection; Formation of development programmes, monitoring and evaluation; Computer models in agriculture: statistical, weather analysis and crop simulation models - concepts, structure, input-output files, limitations, advantages and application for understanding plant processes, sensitivity, verification, calibration and validation; IT application for computation of water and nutrient requirement of crops; Computer-controlled devices (automated systems) for agri-input management; Smartphone mobile apps in agriculture for farm advice, market price, post-harvest management, etc; Geospatial technology, concepts, techniques, components and uses for generating valuable agri-information; Decision support systems, taxonomy, components, framework, classification and applications in agriculture; Agriculture Information/Expert System; Soil Information Systems, etc. for supporting farm decisions; Preparation of contingent crop-planning and crop calendars using IT tools.

# Practical

Study of computer components, accessories; Practice of important DOS commands; Introduction of different operating systems such as windows, Unix, Linux; Creating files and folders; File management; Use of MS-WORD and MS Power point for creating, editing and presenting a scientific document; Handling of tabular data; Animation, video tools, art tool, graphics, template and designs; MS-EXCEL - Creating a spreadsheet, use of statistical tools, writing expressions, creating graphs, analysis of scientific data, handling macros; MS-ACCESS: Creating database, preparing queries and reports, demonstration of agri-information system; Introduction to World Wide Web (WWW) and its components, creation of scientific website, presentation and management agricultural information through web; Introduction of programming languages - Visual Basic, Java, Fortran, C, C++, and their components; Hands-on practice on writing small programmes; Hands-on practice on Crop Simulation Models (CSM); DSSAT/Crop-Info/CropSyst/Wofost; Preparation of input file for CSM and study of model outputs; Computation of water and nutrient requirements of crop using CSM and IT tools; Use of smart phones and other devices in agro-advisory and dissemination of market information; Introduction of generating information; Introduction of generating information; Introduction of generating information important for agriculture; Hands on practice on preparation of Decision Support System.

# **Suggested Readings**

- Gurvinder Singh, Rachhpal Singh & Saluja KK. 2003. Fundamentals of Computer Programming and Information Technology. Kalyani Publishers.
- Harshawardhan P. Bal. 2003. Perl Programming for Bioinformatics. Tata McGraw-Hill Education.
- Kumar A 2015. Computer Basics with Office Automation. IK International Publishing House Pvt Ltd.
- Maidasani D. 2016. Learning Computer Fundamentals, MS Office and Internet & Web Technology. 3<sup>rd</sup> edition, Laxmi Publications.

<b>Course Title</b>	:	Animal Biotechnology
Course No.	:	BTI/VPB/VMC 312
<b>Credit Hours</b>	:	4 (3-0-1)

#### <u>UNIT-I</u>

History and development of animal biotechnology; Basic techniques in animal cell culture: Introduction to embryo biotechnology: oocyte collection and maturation; Sperm preparation; in vitro fertilization; Cryopreservation of oocyte, sperm and embryos; Embryo transfer technology.

#### <u>UNIT II</u>

Breeds of livestock and their characteristics; Marker assisted breeding of livestock; Introduction to animal genomics: RFLP, RAPD, SSRs, QTL, SNP, STR, Mitochondrial DNA polymorphism; Rumen and its environment: Rumen microbes- manipulation of rumen microbes for better utilization of feed; Introduction to nutrigenomics; Milk biome; Manipulation of lactation by biotechnological tools; Application of biotechnology in meat and meat products.

#### <u>UNIT III</u>

Genome and protein based diagnostics of important animal diseases: FMD, brucellosis, PPR, Mastitis, Blue tongue, Newcastle disease; Introduction to vaccinology: live attenuated vaccines, killed vaccines, cell culture based vaccines, recombinant vaccines.

# Practical

Basic cell culture techniques; oocyte aspiration from ovaries; sperm preparation; In vitro fertilization; PCR based detection of animal pathogens; PCR-RFLP; Immuno histochemical localization of protein marker in tissues/cells – meat species identification by PCREDIT

# **Suggested Readings**

Aberle Elton D, Forrest John C, Gerrard David E & Mills Edward W. 2012. *Principles of Meat Science*. 5th Ed. Kendall Hunt Publishing.

- Lawrie & Ledward. Lawrie's. 2006. *Meat Science*. 7th Ed. Woodhead Publishing. Sukumar De. 1997. *Outlines of Dairy Technology*. Oxford University Press-New Delhi.
- Sharma BD. 1999. Meat and Meat Products Technology: Including Poultry Products Technology. Jaypee Bros. Medical Publishers.
- Varnam A & Jane P. 1994. *Milk and Milk Products: Technology, Chemistry and Microbiology*. Sutherland Springer Science & Business Media.

<b>Course Title</b>	:	Immunology
Course No.	:	BTI 311
<b>Credit Hours</b>	:	3 (2-0-1)

## <u>UNIT I</u>

History and scope of immunology; Components of immune system: organs, tissues and cells, Immunoglobulin structure and functions; Molecular organization of immunoglobulins and classes of antibodies; Antibody diversity; antigens, haptens, antigens antibody interactions; Immuno-regulation and tolerance.

#### <u>UNIT II</u>

Allergies and hypersensitive response; Immunodeficiency; Vaccines; Immunological techniques; Immunological application in plant science, monoclonal antibodies and their uses; Molecular diagnostics.

#### Practical

Preparation of buffers and reagents; Precipitation and agglutination test; HA, HI test; Immunoblotting, immunoelectrophoresis and fluorescent antibody test; Enzyme immunoassays including ELISA variants, western blotting; Raising of antisera in laboratory animals; Collection and preservation of antisera – separation, filtration and aliquoting.

## **Suggested Readings**

Murphy K. 2012. Janeway's Immuno Biology. 8th Ed. Garland Science/ Taylor & Francis Group.

Owen JA, Punt J, Kuby J & Sharon A. 2013. Kuby Immunology. 7th Ed. W.H. Freeman

<b>Course Title</b>	:	<b>Molecular Genetics</b>
Course No.	:	BTM 311
<b>Credit Hours</b>	:	2 (2-0-0)

## <u>UNIT I</u>

Structures, properties and modification of DNA; Molecular mechanisms of DNA replication, repair, mutation, and recombination; Centromere and telomere sequences and DNA packaging; Synthesis and processing of RNA and proteins; Regulation of gene expression; Mutations and DNA repair.

# <u>UNIT II</u>

Repetitive DNA sequences and transposable elements; Promoters and their isolation; Transcription factors – their classification and role in gene expression; Epigenetic control of gene expression; Small RNAs, RNA interference and its applications.

# **Suggested Readings**

Allison LA. 2011. Fundamental Molecular Biology. Wiley Global Education.
Brown TA. 1998. Genetics: A Molecular Approach. 3rd Ed. Stanley Thornes.
Lewin B. 2009. Genes 9. Jones & Bartlett Learning.
Tropp BE. 2014. Principles of Molecular Biology. Jones & Bartlett Learning.
Tropp BE. 2012. Molecular Biology Genes to Proteins. 4th Ed. Jones & Bartlett Learning.

<b>Course Title</b>	:	Nanobiotechnology
Course No.	:	BTM 312
<b>Credit Hours</b>	:	2 (2-0-0)

# <u>UNIT I</u>

Introduction to nanotechnology; Concepts and Terminology; Nano-Bio Interface; Biological based Nanosystems, molecular motors, biosensors and other devices.

# <u>UNIT II</u>

Self assembly of molecules for nanotechnology applications; Biomimetics, Biotemplating and *de novo* designed nanostructures and materials; DNA-Nanotechnology; Nanomanipulations, material design, synthesis and their applications.

## **Suggested Readings**

David E. Reisner. 2009. Bionanotechnology: Global Prospects. CRC Press.

Gabor L. Hornyak, John J. Moore, Tibbals HF., Joydeep Dutta. 2008. Fundamentals of Nanotechnology. CRC Press.

Jesus M. de la Fuente, V. Grazu. 2012. Nanobiotechnology: Inorganic nanoparticles Vs Organic nanoparticles. Elsevier.

Yubing Xie. 2012. The Nanobiotechnology Handbook. CRC Press.

<b>Course Title</b>	:	Molecular Marker Technology
Course No.	:	BTF 311
<b>Credit Hours</b>	:	2 (2-0-0)

#### UNIT I

Types of molecular markers- RFLP; PCR based markers like RAPD,SCAR, SSR, STS, CAPS, AFLP, SNP and their variants; Uses of molecular markers: Application as a

genetic tool for genotyping and gene mapping; Mapping populations:  $F_2$ , DH, RILs, NILs; Bulked segregant analysis; Linkage maps; Physical maps (mapping of qualitative genes/QTL); Estimation of genetic similarities and generation of dendrograms.

# <u>UNIT II</u>

Application of molecular markers: Assessing genetic diversity, variety protection; Marker-assisted breeding for accelerated introgression of trait/transgene and quantitative traits; Human and animal health: Association with genetic-based diseases, Paternity determinations; Forensic studies.

# **Suggested Readings**

Huges S. & Moody A. 2007. *PCR: Methods Express*. Royal College of General Practitioners.

<b>Course Title</b>	:	IPR, Biosafety and Bioethics
Course No.	:	BTF 312
<b>Credit Hours</b>	:	2 (2-0-0)

#### <u>UNIT I</u>

Introduction to Intellectual Property, concepts and types; International treaties for protection of IP's; Indian Legislations for the protection of various types of Intellectual Property; Patent search, filing process; Material transfer agreements.

## <u>UNIT II</u>

Biodiversity definition, importance and geographical causes for diversity; Species and population biodiversity, maintenance of ecological biodiversity hot spots in India; Convention on biological diversity; Cartagena Protocol of bio-safety, and risk management for GMO's; Bio-safety guidelines, rules and regulations and regulatory frame work for GMOs in India.

## **Suggested Readings**

Singh BD. 2007. *Biotechnology: Expanding Horizon*. Kalyani Publishers. http://patentoffice.nic.in www.wipo.org www.dbtindia.nic.in www.dbtbiosafety.nic.in

<b>Course Title</b>	:	<b>Genomics and Proteomics</b>
Course No.	:	BTR 311
<b>Credit Hours</b>	:	3 (3-0-0)

### <u>UNIT I</u>

Introduction to Genomics, Functional Genomics and Proteomics; Structural genomics: Classical ways of genome analysis, BAC and YAC libraries; Physical mapping of genomes; Next generation sequencing; Genome analysis and gene annotation; Genome Projects: *E. coli*, Arabidopsis, Bovine, Human; Comparative Genomics: Orthologous and Paralogous sequences, Synteny, Gene Order, Phylogenetic footprinting.

# <u>UNIT II</u>

Functional genomics: Differential gene expression techniques: ESTs, cDNA-AFLP, microarray, Differential display, SAGE, RNAseq, Real time PCREDIT

# <u>UNIT III</u>

Introduction to proteomics; Analysis of proteome: Native PAGE, SDS PAGE, 2D PAGE; Edmann Degradation; Chromatographic techniques: HPLC, GC, Mass Spectrometry: MALDI-TOF, LC-MS; Post Translational modifications.

#### **Suggested Readings**

Branden C & Tooze J. 1999. Introduction to Protein Structure. 2<sup>nd</sup> Ed. Garland Science.

- Connor DO & Hames BD. 2007. *Proteomics: Methods Express*. Royal College of General Practitioners.
- Pennington S R &Dunn M J. 2001. Proteomics from protein sequence to function. BIOS Scientific Publishers Ltd.

Singer M & Berg P. 1991. *Genes & Genome*. University Science Books. Tropp BE. 2012. *Molecular Biology Genes to Proteins*. 4th Ed. Jones & Bartlett Learning

<b>Course Title</b>	:	<b>Enzymology and Enzyme Technologies</b>
Course No.	:	BTB 311
<b>Credit Hours</b>	:	3 (2-0-1)

# <u>UNIT I</u>

Classification and nomenclature of enzymes; General characteristics of enzymes, active site, cofactors, prosthetic groups; Metalloenzymes; Isolation, purification, characterization and assays of enzyme and international units; Criteria for purity.

# <u>UNIT II</u>

Enzyme kinetics: effect of pH, temperature, determination of Km and Vmax; Regulation of enzyme activity; Enzyme inhibition: competitive, non-competitive and uncompetitive; Isoenzymes, schizomers and isoschizomers; Ribozymes; Immobilization of enzymes; Applications of enzymes: biotechnology, industry, environment, agriculture, food and medicine.

# Practical

Isolation, purification and assay of enzymes;Determination of optimum pH and optimum t;emperature of enzymes; Thermostability of enzymes; Activators and inhibitors of enzyme catalysis; Determination of kinetic parameters of enzymes; Immobilization of enzymes; Isoenzymes analysis.

# **Suggested Readings**

Bisswanger H. 2011. Practical Enzymology. 2nd Ed. Wiley-Blackwell.

Cook PF & Cleland WW. 2007. *Enzyme Kinetics and Mechanism*. Garland Publishing Inc.

Cornish-Bowden A. 2012. Fundamentals of Enzyme Kinetics. 4th Ed. Wiley-Blackwell. Price NC & Stevens L. 1999. Fundamentals of Enzymology: Cell and Molecular Biology of Catalytic Proteins. 3rd Ed. Oxford University Press.

# **SEMESTER-VI**

<b>Course Title</b>	:	<b>Biostatistics</b>
Course No.	:	BAS 321
<b>Credit Hours</b>	:	3 (2-0-1)

# <u>Unit I</u>

Random variables: expected value and its variance; probability distribution of random variables; Conditional probability; Baye's theorem and its applications; Introduction to Uniform, Binomial, Poisson, Normal, Exponential and Gamma probability distributions.

# <u>Unit II</u>

Random mating populations, Hardy-Weinberg Law; Introduction to Poisson process and Markov chains: Transition probability matrix, n-step transition probabilities, steady state. Random walk models; Sensitivity and specificity.

# <u>Unit III</u>

Chi-square test: testing heterogeneity, use in genetic experiment, detection of linkage, linkage ratios and its estimation; Analysis of variance: One-way and two-way classification with interaction; Analysis of covariance; Incomplete block designs; Estimation and significance of genotypic and phenotypic variation.

# Practical

Expected value and variance of discrete and continuous distributions; Uniform, Binomial, Poisson, Normal, Exponential and Gamma Probability distributions; Hardy-Weinberg Law; Construction of transition probability matrix in Markov Chains; Calculation of sensitivity and specificity; Detection and linkage using Chi-square test; One-way and two-way analysis of variance; Analysis of covariance; Incomplete block designs; Testing of heritability.

# **Suggested Readings**

Biswal PC. 2009. Probability and Statistics. PHI Learning Pvt. Ltd.

Kaps M. & Lamberson W. 2007. Biostatistics for Animal Science. CABI Publishing.

Narayan P, Bhatia VK & Malhotra PK. 1989. *Handbook of Statistical Genetics*. Indian Agricultural Statistics Research Institute, New Delhi, India.

Pal N. & Sahadeb Sarkar. 2009. *Statistics – Concepts and Applications*. 2<sup>nd</sup> Ed. PHI Learning Pvt. Ltd.

<b>Course Title</b>	:	<b>Computational Biology</b>
Course No.	:	<b>BTI/BTR 321</b>
<b>Credit Hours</b>	:	3 (2-0-1)

<u>UNIT I</u>

Introduction to computational biology; Web based servers and software for genome analysis: Ensembl, UCSC genome browser, MUMMER, BLASTZ; Sequence submission.

# UNIT II

Protein interaction databases: BIND, DIP, GRID, STRING, PRIDE; Principles of Protein structure prediction; Fold Recognition (threading); Homology modeling; SCOP, CATH, PDB, PROSITE, PFAM; Methods for comparison of 3D structures of proteins.

#### <u>UNIT III</u>

Phylogenetic analysis: Evolutionary models, tree construction methods, statistical evaluation of tree methods; PHYLIP, dendroscope, MEGA; DNA barcoding database-BOLD.

# Practical

Application of Genome browsers in genomic research; Exploring protein-protein interaction databases; Working with protein structural classification databases; SNP and SSR identification tools; PHYLIP.

## **Suggested Readings**

Creighton TE. 1993. Proteins: Structures and Molecular Properties 2<sup>nd</sup> Edition. W.H Freeman.

DovStekel. 2003. Microarray Bioinformatics. 1st Ed. Cambridge University Press.

- Mount D. 2001. *Bioinformatics: Sequence and Genome Analysis*, 2<sup>nd</sup> Ed. Cold Spring Harbor Laboratory Press.
- Malcolm Campbell A. & Laurie J. Heyer. 2007. Discovering Genomics, Proteomics and Bioinformatics. 2<sup>nd</sup> Ed. Benjamin Cummings.
- Setubal Joao & Meidanis Joao. 2004. *Introduction to Computational Molecular Biology*, PWS Publishing Company.

# **Elective I. Plant Biotechnology**

<b>Course Title</b>	:	Plant Tissue Culture and its Applications
Course No.	:	BTF 321
<b>Credit Hours</b>	:	3 (2-0-1)

# Theory

# <u>UNIT I</u>

Historical benchmarks of plant cell and tissue culture; Culture media components and modifications; Sterilization techniques; Various types of culture: callus, suspension, nurse, root, meristem; *In vitro* differentiation: Organogenesis and somatic embryogenesis; Plant growth regulators: mode of action, effects on *in vitro* culture and regeneration.

#### <u>UNIT II</u>

Applications: Micropropagation; Anther and microspore culture; Somaclonal variation; *In vitro* mutagenesis; Production of secondary metabolites; Synthetic seeds; *In vitro* fertilization; Embryo rescue in wide hybridization; Endosperm culture; Protoplast isolation, culture and regeneration; Somatic hybridization: cybrids, asymmetric hybrids; *In vitro* germplasm conservation.

# Practical

Establishment of callus/ cell suspension cultures; Induction of plant regeneration; Micropropagation – Explant establishment, shoot multiplication, root induction, Hardening and transfer to soil; Monitoring of growth and differentiation of cells, Seed/Embryo culture; Ovary culture, Anther /pollen culture, Suspension cultures and production of secondary metabolites.

# **Suggested Readings**

Bhojwani SS & Razdan MK. 1996. Plant Tissue Culture: Theory and Practice. Elsevier.

- Debergh PC & Zimmerman RH. 1991. *Micropropagation: Technology and Application*. Kluwer Academic.
- Dixon RA & Gonzales RA. 2003. *Plant Cell Culture: A Practical Approach*. Oxford University press.

George EF, Hall MA & Klerk GJD. 2007. *Plant Propagation by Tissue Culture*. 3<sup>rd</sup> Ed. Volume 1. Springer Science & Business Media.

<b>Course Title</b>	:Princ	iples and Applications of Plant Genetic Transformation
Course No.	:	BTM 321
<b>Credit Hours</b>	:	3 (2-0-1)

# <u>UNIT I</u>

Gene transfer methods: Direct and Indirect; Marker free transformation; *In planta* transformation; Vectors for plant transformation, molecular characterization of transgenic plants using PCR, real time PCR, Southern, Northern and western analysis; Bioassays with transgenic plants; Evaluation and selection of transgenic events for target trait.

# <u>UNIT II</u>

Genetic engineering of crop plants for useful traits: Over expression, inducible, tissue specific and gene silencing systems; Biosafety concerns and regulatory mechanisms; Commercialization of transgenic products, GMO's, transgenic plants for the production of biopharmaceuticals; Molecular farming of plants for applications in medicine systems, heterologous protein production in transgenic plants; Successful case studies.

# Practical

Gene isolation and gene cloning; Gene constructs and their maintenance; *Agrobacterium* mediated genetic transformation; Particle gun mediated genetic transformation. Histochemical GUS assays; PCR screening of putative transgenic plants; Raisingtransgenic under containment and field conditions.

# **Suggested Readings**

Bhojwani SS & Dantu PK. 2013. Plant Tissue Culture: An Introductory Text. Springer

Brown TA. 2007. Gene Cloning & DNA Analaysis: An Introduction. 6 <sup>th</sup> Ed. Wiley	y-
Blackwell Publishing.	
Grierson D. 2012. Plant Genetic Engineering. Springer Netherlands.	
Lal R & Lal S. 1990. Crop Improvement Utilizing Biotechnology. CRC Press.	
Primose SB & Twyman RM. 2006. Principles of Gene Manipulation and Genomics. 7	7 <sup>th</sup>
Ed. Wiley-Blackwell Publishing.	

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<b>Course Title</b>	:	<b>Epigenetics and Gene regulation</b>
Course No.	:	BTM 322
<b>Credit Hours</b>	:	3 (2-0-1)

#### <u>UNIT I</u>

DNA methylation and histone modifications: DNA methylases, methyl binding proteins and histone modifiers; Epigenetic changes in response to external stimuli leading to changes in gene regulation; Role of DNA methylation in plant development: mutant case studies.

#### <u>UNIT II</u>

Introduction to small RNAs: History, biogenesis; *In silico* predictions, target gene identification, methylation of heterochromatin by het associated siRNAs; Gene regulation by small RNA Other classes of siRNAs; Role in epigenetics; Jacob Monod model; RNA editing, Genome imprinting.

# Practical

*In silico* study of structural components of histone modifiers and DNA methylases of model plants; *In silico* prediction of siRNAs and miRNAs; Small RNAs electrophoresis using PAGE; Blotting of small RNAs on nylon membrane; miRNA target finding; Detection of small RNAs using fluorescent labelled probes; Bisulphite sequencing for methylation; qRT-PCR for quantitative analysis of small RNAs in developmental phases.

# **Suggested Readings**

Green & Sambrook. 2014. *Molecular Cloning: A Laboratory Manual*. 4<sup>th</sup> Ed. Vol I, II & III Cold Spring Harbor Laboratory Press.

Mohanpuria P, Kumar V, Mahajan M, Mohammad H & Yadav SK. 2010. Gene Silencing: Theory, Techniques and Applications: Genetics-Research and Issues. Nova Science Publishers.

<b>Course Title</b>	:	<b>Applications of Genomics and Proteomics</b>
Course No.	:	BTR 322
<b>Credit Hours</b>	:	3 (2-0-1)

#### <u>UNIT I</u>

Structure of genomes: *Arabidopsis*, rice, tomato, pigeon pea, wheat; DNA chips and their use in transcriptome analysis; Mutants and RNAi in functional genomics; Site directed mutagenesis; Transposon tagging; Transient gene expression: VIGS and FACS based, targeted genome editing technologies.

# <u>UNIT II</u>

Bio-informatics in proteomics: Protein 3D structure modelling (Homology modelling and crystallography); Proteome analysis; Protein- protein interaction: FRET, yeast two hybrid and co-immunoprecipitation. Applications of genomics and proteomics in agriculture, human health and industry. Metabolomics and ionomics for elucidating metabolic pathways.

# Practical

SDS\_PAGE; 2D Electrophoresis; Protein characterization through HPLC; Specialized crop based genomic resourses: TAIR, Gramene, Graingenes, Maizedb, Phytozome, Cerealdb, Citrusdb; miRbase.

# **Suggested Readings**

Connor DO & Hames BD. 2007. Proteomics: Methods Express. Royal College of General Practitioners.

Pennington S R, Dunn M J. 2001. Proteomics from protein sequence to function. BIOS Scientific Publishers Ltd.

Singer M & Berg P. 1991. Genes & Genome. University Science Books.

- Tropp BE. 2012. *Molecular Biology Genes to Proteins*. 4th Ed. Jones & Bartlett Learning.
- Verma PS & Agarwal VK. 2014. Cell Biology, Genetics, Molecular Biology, Evolution and Ecology. S. Chand & Company Pvt. Ltd.

<b>Course Title</b>	:	Molecular Breeding in Field Crops
Course No.	:	GPB 322
<b>Credit Hours</b>	:	3 (2-0-1)

#### <u>UNIT I</u>

Principles of plant breeding; Breeding methods for self and cross pollinated crops; Heterosis breeding; Limitations of conventional breeding; Development of specific mapping populations.

#### <u>UNIT II</u>

QTL mapping using structured populations; Fine mapping of genes/QTL; Map based gene/QTL isolation and development of gene based markers.

# <u>UNIT III</u>

Marker assisted selection (MAS): Foreground and background selection; MAS for majorand minor genes, Marker assisted pyramiding, Marker assisted recurrent selection; Transgenic breeding; MAS for specific traits with examples; Commercial applications of MAS.

# Practical

Working on some genotyping and phenotyping datasets for Linkage mapping using softwares such as Mapmaker, MapDisto and QTL mapping softwares such as WinQTL cartographer; Use of gene based and closely linked markers for foreground selection for target traits in target crops; Marker assisted detection of the transgene.

## **Suggested Readings**

Nagat T, Lorz H & Widholm JM. 2008. *Biotechnology in Agriculture and Forestry*. Springer.

Trivedi PC. 2000. Plant Biotechnology: Recent Advances. Panima Publishers.

<b>Course Title</b>	:Molecular Breeding of Horticultural Crops and Forest Tree	
Course No.	:	HOR 322
<b>Credit Hours</b>	:	3 (2-0-1)

#### <u>UNIT I</u>

Reproductive biology of major fruit and forest crops; Basic methods of fruit crop improvement; Target traits in major fruit crops; Limitations of fruit crop breeding; Breeding methods of self and cross pollinated vegetable crops; Breeding of commercial flower crops.

# <u>UNIT II</u>

Molecular markers for germplasm characterization and genetic diversity analysis; Pseudo test cross mapping strategy in fruit crops; Molecular mapping in vegetable crops; Marker assisted breeding in horticultural crops and forest plants; Micropropagation for variety dissemination; Mutation breeding and characterization of mutants; Genomic resources for marker development; Transgenic approaches with tree crops and utility.

## Practical

Modifications in DNA extraction methods for horticultural and forest crops; Agarose gel electrophoresis, and DNA quantification; Map maker; Diversity analysis using UPGMA; Identifying repeat sequences using MISA; Standard Gene cloning methods including construct making with the use of Restriction enzymes; DNA ligases and standard molecular approaches.

# **Suggested Readings**

J S Bal. 2013. Fruit Growing. Kalyani Publishers.

Kumar N. 2006. Breeding of Horticultural crops: Principles and Practices. New India Publishing Aagency.

K. L. Chada. 2012. Handbook of Horticulture. ICAR.

Kumar J. Prasad. 2010. Handbook of Fruit Production. Agrobios.

Schnell RJ & Priyadarshan PM. 2012. Genomics of Tree Crops. Springer.

Singh Jitender. 2014. Basic Horticulture. Kalyani Publishers.

Singh Ranjit. 2012. Fruits. National Book Trust.

Spangenberg G. 2001. *Molecular Breeding of Forage Crops*. Kluwer Academic Publishers.

Victor Ray Garden, Frederick Charles Bnaford, Herry & Daggett HoDlor Ir. 1992.

Fundamentals of Fruit Production. Mc Graw Book Company.

# **Elective II. Animal Biotechnology**

<b>Course Title</b>	:	<b>Animal Genomics</b>
Course No.	:	VPB 321
<b>Credit Hours</b>	:	3 (2-0-1)

#### UNIT I

Genome organization in eukaryotes; Satellite DNA: VNTRs & families, LINE & SINE; Sex determination: Chomosomal basis of sex determination, Molecular markers for sexdetermination, environmental sex determination; Chromosomal aberrations: Euploidy, Chromosomal Non-disjunction and Aneuploidy, Polyploidy, Induced Polyploidy, Syndromes, Structural aberrations, Robertsonian Translocations, Position Effect, Chromosomal Mosaics, Chromosomal aberrations and evolution.

# <u>UNIT II</u>

Molecular Markers: Markers, Genetic Markers: RAPD, STR, DNA fingerprinting, SSCP, RFLP, SNP, EST; SNP Analysis; karyotyping, Somatic cell hybridization; Radiation hybrid maps; FISH technique; Major Histocompatibility Complex: Concept and its relevance in disease resistance & immune response; Quantitative trait Loci; Marker Assisted Selection: Concept, Linkage Equilibrium, Application in Animal Sciences; Genomic Selection: Concept, Linkage Disequilibrium, Methodologies of economic Selection; Mitochondrial DNA analysis and its application in livestock; Applying DNA markers for breed characterization.

# Practical

Extraction of genomic DNA from peripheral blood; Analysis of DNA by agarose or polyacrylamide gel electrophoresis; Checking the quality & quantity of genomic DNA; Restriction digestion & analysis; Sanger Sequencing data analysis; Extraction of mitochondrial DNA; Extraction of RNA from PBMC; Quality checking of total RNA; cDNA synthesis.

#### **Suggested Readings**

Brown TA. 2006. Genomes. 5<sup>th</sup> Ed. Wiley-Blackwell.

Dale JW, Schantz MV & Plant N. 2012. From Genes to Genomes: Concepts and Applications of DNA Technology. John Wiley & Sons.

Green & Sambrook. 2014. *Molecular Cloning: A Laboratory Manual*. 4<sup>th</sup> Ed. Vol I, II & III. Cold Spring.

Reece RJ. 2004. Analysis of Genes & Genomes. Wiley.

<b>Course Title</b>	:	Embryo Transfer Technologies
Course No.	:	VPB/VGO 322
<b>Credit Hours</b>	:	3 (2-0-1)

#### <u>UNIT I</u>

History, advantages, limitations and scope of embryo transfer technology; Estrus cycle and its detection in animals; Methodology of super ovulation; Ovum pick up (OPU);

Preparation of sperm for *in vitro* fertilization (IVF); Embryo grading and culture; Micromanipulation and immuno-modulation for enhancement of fecundity.

# UNIT II

Different methods of gene transfer and their limitations; embryo splitting; embryo sexing by different methods; production of transgenic livestock by nuclear transfer and its application; regulatory issues (social, ethical, religious and environmental); Cloning of domestic animals; Conservation of endangered species; Characterization of embryonic stem cells and applications.

# Practical

Demonstration of estrus detection methods; Estrus synchronization; Superovulation; Oocyte collection from slaughterhouse ovaries; Grading of oocytes from slaughterhouse ovaries; collection and preparation of semen samples; *In vitro* fertilization; Collection of embryos using non-surgical procedures; Grading and culture of embryos; Embryo sexing by different methods; Embryo splitting; Embryo freezing.

## **Suggested Readings**

Gordon I. 2004. Reproductive Technologies in Farm Animals. CABI. Hafez ESE. 2000. Reproduction in Farm Animals. Lippincott, Williams & Wilkins.

<b>Course Title</b>	:	Principles and Procedures of Animal Cell Culture
Course No.	:	BTI/VPB/VMC 323
<b>Credit Hours</b>	:	3 (2-0-1)

#### <u>UNIT I</u>

History, importance and development of animal cell culture techniques; Basic requirements for animal cell culture; Sterilization procedures for cell culture work; Different types of cell culture media, growth supplements, serum free media and other cell culture reagents.

#### <u>UNIT II</u>

Different cell culture techniques including primary and secondary cultures; continuous cell lines, suspension culture, organ culture etc; Commonly used animal cell lines: CHO, HeLa, BHK-21, VERO, Sf9, C636; Their origin and characteristic, growth kinetics of cells in culture, differentiation of cells; Characterization and maintenance of cell lines; Applications of animal cell cultures.

#### <u>UNIT III</u>

Cryopreservation and revival of cells; Hybridoma technology; Scaling up methods; bioreactors; Overview of insect cell culture; Stem cell culture and its application; Common cell culture contaminants and their management.

# Practical

Basic equipments used in animal cell culture laboratories; Washing, packing and sterilization of glass and plastic wares for cell culture; Preparation of media and reagents for cell culture; Primary culture technique of chicken embryo fibroblast; Culture and sub-culturing of continuous cell lines; Viability assay by trypan blue dye exclusion method; Isolation and cultivation of lymphocytes; Cryopreservation of primary cultures and cell lines; Cytopathic effect of viruses on cultured mammalian cells.

# **Suggested Readings**

Butler M. 2003. Animal Cell culture & Technology. Garland Science.
Freshney RI. 2011. Culture of Animal Cells: A manual of basic technique and specialized applications. 6<sup>th</sup> Ed. John Wiley & Sons.

<b>Course Title</b>	:	<b>Transgenic Animal Production</b>
Course No.	:	<b>BTI/VPB 324</b>
<b>Credit Hours</b>	:	3 (3-0-0)

## <u>UNIT I</u>

History of transgenesis; Isolation of gene, preparation of gene construct; Methods of transgenic animal production: Calcium chloride mediated transfection, lipofection, electroporation, microinjection, nanodelivery.

#### <u>UNIT II</u>

Production of gene knockouts: cre-lox, zinc finger nucleases; CRISPR; TALENs; Production of chimeric animals; gene silencing by lentivirus system.

## <u>UNIT III</u>

Stem cell technology: Isolation and characterization of stem cell lines from different sources: embryo, mesenchymal, induced pluripotent stem cell; Introduction to animal cloning; Application of stem cells in transgenesis and animal cloning.

#### <u>UNIT IV</u>

Fundamental assays of transgenic products: confirmation of integration of transgene; Validation of transgenic products like isolation of transgenic protein from milk and characterization; Application of transgenics in production of disease resistance models and carcinogenesis. Regulatory issues associated with transgenic animal production.

#### **Suggested Readings**

Ramadass P. 2008. Animal Biotechnology: Recent Concepts and Developments. MJP Publishers.

Ranga MM. 2007. Animal Biotechnology. Agrobios.

Singh BD. 2010. Biotechnology expanding Horizons. Kalyani Publishers.

Singh B. Gautam SK & Chauhan MS. 2014. Textbook of Animal Biotechnology. The Energy and Resources Institute, TERI.

<b>Course Title</b>	:	<b>Molecular Diagnostics</b>
Course No.	:	BTI/VPB 325
<b>Credit Hours</b>	:	3 (2-0-1)

## <u>UNIT I</u>

Principle and applications of molecular diagnostic tests; Nucleic acid based diagnostics for detection of pathogenic organisms: Application of restriction endonuclease analysis for identification of pathogens; Polymerase chain reaction (PCR) and its variants; Reverse transcriptase polymerase chain reaction (RT PCR); isothermal amplification (LAMP); LCR, nucleic acid sequence-based amplification (NASBA); Real-Time PCR; DNA Probes; Southern blotting; Northern blotting; Protein based assays: SDS-PAGE, Western Blot, Dot-blot, ELISA and lateral flow device.

#### <u>UNIT II</u>

Advantages of Molecular diagnostics over conventional diagnostics; serodiagnostics; DNA array technology; Protein array; tissue array; Biosensors and nanotechnology; Development and validation of diagnostic tests.

## Practical

Preparations of buffers and reagents; Collection of clinical and environmental samples for molecular detection of pathogens (bacteria/virus); Extraction of nucleic acids (DNA & RNA) from the clinical specimens; Restriction endonuclease digestion and analysis using agarose gel electrophoresis; Polymerase chain reaction for detection of pathogens in blood and animal tissues; RT-PCR for detection of RNA viruses; PCR based detection of meatadulteration in processed and unprocessed meats; PCR based detection of pathogens in milk, eggs and meat; Lateral flow assay; ELISA.

#### **Suggested Readings**

Debnath M, Prasad GBKS & Bisen PS. 2010. *Molecular Diagnostics: Promises and Possibilities*. Springer Science & Business Media.

Singh BD. 2010. Biotechnology expanding Horizons. Kalyani Publishers.

Viljoen, GJ, Nel LH & Crowther JR. 2005. *Molecular Diagnostic PCR Handbook*. Springer Science & Business Media.

Wilson K & Walker J. 2010. Principles and Techniques of Biochemistry and Molecular Biology. Cambridge University Press.

<b>Course Title</b>	:	<b>Molecular Virology and Vaccine Production</b>
Course No.	:	BTI/VMC 326
<b>Credit Hours</b>	:	3 (2-0-1)

# <u>UNIT I</u>

Properties of viruses; Classification of viruses; Virus replication; Cell transformations, Cultivation of viruses, assay techniques for detection/quantification; Important Animal viruses; Virus-Host interactions; Viral infections; Immune responses to viruses: Interferon and other cytokines; Bio-safety and bio-security principles.

#### <u>UNIT II</u>

Properties of an ideal vaccine; Classification of vaccines; Methods of inactivation and attenuation of viruses; New generation vaccines: subunit, synthetic, rDNA, marker and edible; Adjuvants and vaccine delivery systems; Novel immunomodulators and vaccine

delivery using nanotechnology; Vaccine preparation: Stabilizers, preservatives and vehicles; Quality control and testing of vaccines; Sero-surveillance and sero-monitoring.

# Practical

Processing of clinical specimens for isolation of viruses; Cultivation of viruses in cell cultures and embryonated eggs; Harvesting of virus; Study of cytopathic effects; Titration of virus and estimation of TCID<sub>50</sub>;Haemagglutination and Haemagglutination Inhibition test; Detection of virus by SNT, AGID and ELISA.

# **Suggested Readings**

John Carter J & Saunders V. 2007. *Virology: Principles and Applications*. 2<sup>nd</sup> Ed. Wiley. Morrow WJW, Sheikh NA, Schmidt CS, Davies DH. 2012. *Vaccinology: Principles and Practice*. John Wiley & Sons.

Sharma S & Adlakha S. 1996. *Textbook of Veterinary Microbiology*. Vikas Publishing House Pvt Ltd.

Stephenson J & Warnes R. 1998. *Diagnostic Virology Protocols*. Springer Science & Business Media.

**Elective III. Microbial and Environmental Biotechnology** 

<b>Course Title</b>	:	Microbial Biotechnology
Course No.	:	BTP 321
<b>Credit Hours</b>	:	3 (2-0-1)

#### <u>UNIT I</u>

Microbial biotechnology, scope and techniques; Industrially important microorganisms; Gene transfer mechanisms in microbes: Transformation, transduction, conjugation and recombination; Genetic variability in microorganisms; Biotechnological tools to improve the microbial strains with respect to industry and agriculture.

#### <u>UNIT II</u>

Biotransformation and biodegradation of pollutants, biodegradation of lignocelluloses and agricultural residues; Biotechnological treatment of waste water, sewage and sludge; Industrial production of alcohols, ethanol, acids (citric acid, acetic acid), solvents (glycerols, acetone, butanol), antibiotics (penicillin, streptomycine, tetracycline), amino acids (lysine, glutamic acid), single cell proteins; Recombinant and synthetic vaccines.

#### Practical

Isolation and preservation of industrially important microorganisms; Microbial fermentation, production of proteins and enzymes using bacteria, yeast and fungus; Microbial biomass production, utilization of plant biomass by recombinant microorganisms; Production of secondary metabolites from microbes.

# **Suggested Readings**

Glaze AN & Nikaido H. 2007. *Microbial Biotechnology: Fundamentals of Applied Microbiology*. 2<sup>nd</sup> Ed. Cambridge University Press.

Mohapatra PK. 2006. *Text Book of Environmental Biotechnology*. International Publishing House Pvt. Ltd.

<b>Course Title</b>	:	Green Biotechnology
Course No.	:	<b>BTP/BTR 324</b>
<b>Credit Hours</b>	:	3 (2-0-1)

## <u>UNIT I</u>

Green biotechnology: Definition, concept and implication; Bio-fertilizers and biopesticides; Plant growth promoting rhizobacteria; Production of biofuels, biodiesel and bioethanol; Biomass enhancement through biotechnological interventions; Generation of alternate fuels in plants; Identification and manipulation of micro-organisms for biodegradation of plastics and polymers; GMOs for bioremediation and phytoremediation, their roles; Strategies for detection and control of soil, air and water pollutants.

## <u>UNIT II</u>

Carbon sequestration; Methanogenic microbes for methane reduction; Microbes for phytic acid degaradation; Genetic Engineering for increasing crop productivity by manipulation of photosynthesis, nitrogen fixation and nutrient uptake efficiency; Marker-free transgenic development strategies; Development of disease resistant and pest resistant crops through biotechnological tools.

# Practical

Identification and efficiency assays of micro-organisms for biodegradation and bioremediation; Isolation of *Bacillus thurigenesis* and plant growth promoting rhizobacteria; Production of biofertilizers, biopesticides and biofuel; Assays for removal of oil spillage.

#### Suggested Readings

Kirkosyan A & Kaufman PB. 2009. *Recent Advances in Plant Biotechnology*. Springer. Kumar A. 2004. *Environmental Biotechnology*. Daya Publishing House. Murray DC. 2011. *Green Biotechnology*. Dominant Publishers and Distributors.

<b>Course Title</b>	:	Molecular Ecology and Evolution
Course No.	:	BTC 321
<b>Credit Hours</b>	:	3 (3-0-0)

# <u>UNIT I</u>

Molecular Evolution: Concept, molecular divergence and molecular clocks; Speciation and domestication; Evolution of earth and earlier life forms; Primitive organisms, their metabolic strategies and molecular coding; New approaches to taxonomical classification including ribotypeing, Ribosomal RNA sequencing; Molecular tools in phylogeny, classification and identification.

# <u>UNIT II</u>

Protein and nucleotide sequence analysis; Origin of new genes and proteins; Gene duplication and divergence; Genome evolution, components of genomes, whole genome duplications, chromosome rearrangements and repetitive sequence evolution.

#### <u>UNIT III</u>

Application of molecular genetics and genomics to ecology and evolution; Assessment of genetic diversity, phylogeny, inbreeding, quantitative traits using molecular tools; Mutations; Regulations of gene expression.

#### **Suggested Readings**

Beebee T & Rowe G. 2008. An Introduction to Molecular Ecology. 2nd Ed. Oxford University Press.

Brown TA. 2007. *Genome 3*. Garlan Science Publishing. Carvalho GR. 2002. *Advances in Molecular Ecology*. IOS Press Netherland.

<b>Course Title</b>	:	Molecular Pharming and Biopharmaceuticals
Course No.	:	BTI/VPT 327
<b>Credit Hours</b>	:	3 (2-0-1)

# <u>UNIT I</u>

Concept of molecular pharming and production of biopharmaceuticals; Mammalian cell culture manufacturing and microbial fermentation; Fermentation and cell culture processing; Protein purification and processing; Industrial fermentation: batch and continuous cultures, production of biopharmaceuticals, immobilization techniques.

### <u>UNIT II</u>

Biopharmaceutical analytical techniques; Biopharma drug discovery and development; production of specific vaccines and therapeutic proteins.

#### Practical

Isolation & purification of proteins from microbes and plants; Production of recombinant proteins in prokaryotes; Analysis of proteins by one and two dimensional gel electrophoresis; Affinity chromatography; Immunoblotting; Cell culture and immobilization techniques. Visit to biopharmaceutical industry.

# **Suggested Readings**

Brown TA. 2010. Gene Cloning and DNA analysis: An Introduction. 6<sup>th</sup> Ed. Wiley-Blackwell Publishing.

Kirkosyan A & Kaufman PB. 2009. Recent Advances in Plant Biotechnology. Springer.

Primrose SB & Twyman RM. 2013. Principles of Gene Manipulation and Genomics. John Wiley & Sons.

<b>Course Title</b>	:	Food Biotechnology
Course No.	:	BTO/AGE 321
<b>Credit Hours</b>	:	3 (2-0-1)

# <u>UNIT I</u>

Food Biotechnology: Introduction, history and importance; Applications of biotechnology in food processing: Recent developments, risk factors and safety regulations; Food spoilage and preservation process; Food and beverage fermentation: Alcoholic and non alcoholic beverages, food additives and supplements.

# <u>UNIT II</u>

Industrial use of micro organisms; Commercially exploited microbes: *Saccharomyces, Lactobacillus, Penecillium, Acetobactor, Bifidobacterium, Lactococcus* and *Streptococcus*; Dairy fermentation and fermented products; Prebiotics and probiotics; Genetic engineering for food quality and shelf life improvement; Bioactive peptides; Labelling of GM foods.

## Practical

Isolation, culture and maintenance of biotechnologically important micro-organisms; Use of laboratory and industrial scale shakers; Batch and continuous cultures; Use of fermentors; Detection of pathogens in food and feed; Detection of GM food; Visit to food processing industry.

# **Suggested Readings**

- Hui YH & Khachatourians GG. 1995. Food Biotechnology: Microorganisms. Wiley-VCH
- Shetty K, Paliyath G, Pometto A. & Levin RE. 2006. Food Biotechnology. 2<sup>nd</sup> Ed. CRC Press.

<b>Course Title</b>	:	<b>Bio-prospecting of Molecules and Genes</b>
Course No.	:	BTO 322
<b>Credit Hours</b>	:	3 (3-0-0)

## <u>UNIT I</u>

Concepts and practices of bioprospecting; Traditional and modern bioprospecting; Gene prospecting; Isolation, synthesis and purification of new bioactive chemicals for laboratory. clinical and field trials; Intellectual property rights, mechanisms and the legal framework; Patenting of new genes and/or bioactive principles with novel antibiotic, insecticidal or anti-tumour properties.

### <u>UNIT II</u>

Principles of the Convention on Biological Diversity, biodiversity conservation and biotechnology; Development and management of biological, ecological, taxonomic, and related systematic information on living species and systems.

# <u>UNIT III</u>

Bioprospecting of microorganisms and their components; Bioprospecting of biodiversity for new medicines: Identification and collection of material by random and traditional (medicinal) approaches; Screening for particular bio-activities; Elucidation of novel molecular form, process technology; Development of techniques for large scale industrial production of the final bioactive product and its market availability and accessibility to the public.

# **Suggested Readings**

Mohapatra PK. 2006. *Text Book of Environmental Biotechnology*. International Publishing House Pvt. Ltd.

Sharma PD. 2012. Ecology and Environment. 11th Ed. Rastogi Publications.

# **Elective IV. Bioinformatics**

<b>Course Title</b>	:	<b>Programming for Bioinformatics</b>
Course No.	:	BAS 322
<b>Credit Hours</b>	:	4 (2-0-2)

#### <u>UNIT I</u>

Introduction: Operating systems, programming concepts, algorithms, flow chart, programming languages, compiler and interpreter; Computer number format: Decimal, Binary, Octal and Hexadecimal.

## <u>UNIT II</u>

C-Language: History, constant, variables and identifiers, character set, logical and relational operators, data input and output concepts; Decision making: if statement, ifelse statement, for loop, while loop and do-while loop; Arrays and functions, file handling; Programs related to arithmetic operations, arrays and file handling in C.

# Practical

#### <u>UNIT I</u>

PERL-Language: Introduction, variables, arrays, string, hash, subroutines, file handling, conditional blocks, loops string operators and manipulators, pattern matching and regular expressions in PERL; Sequence handling in PERL demonstrating string, array and hash.

#### <u>UNIT II</u>

Shell Programming: Concepts and types of UNIX shell, Linux variables, if statements, control and iteration, arithmetic operations, concepts of awk, grep and sed; Sequence manipulations using shell scripting.

## **Suggested Readings**

Balagurusamy. 2008. *Programming in ANSI C*. Tata McGraw-Hill Education. James Tisdall. 2003. *Mastering Perl for Bioinformatics*. O'Reilly Media.

Tom Christiansen, Brian D Foy, Larry Wall & Jon Orwant. 2012. *Programming Perl.* 4<sup>th</sup>Ed. O'Reilly Media. Kanetkar Yashavant. 2013. *Let Us C.* BPB Publications.

<b>Course Title</b>	:	<b>Computational Methods for Data Analysis</b>
Course No.	:	BAS 323
<b>Credit Hours</b>	:	2 (1-0-1)

# <u>UNIT I</u>

Introduction to UNIX/LINUX operating system; Knowledge discovery and data mining techniques; Machine learning and pattern recognitions, hidden markov models; Artificial neural networks, Support vector machines.

### <u>UNIT II</u>

Principal component analysis, ANOVA; AMOVA and different clustering methods; Gene Prediction algorithms and Phylogeny algorithms; Basics of R statistical package.

# Practical

Gene prediction: FGENESH; R statistical package installation and configuration, GUI for R: R-commander, R-studio, RKWard; Analysis of gene expression using R; GNU PSPP, Scilab, QtiPlot.

# **Suggested Readings**

Gareth James, Daniela Witten, Trevor Hastie & Robert Tibshirani. 2013. An Introduction to Statistical Learning: with Applications in R. Springer
 Mathur K Sunil. 2010. Statistical Bioinformatics with R. Elsevier.

<b>Course Title</b>	:	<b>Bioinformatics Tools and Biological databases</b>
Course No.	:	<b>BTI/BTR 328</b>
<b>Credit Hours</b>	:	3 (2-0-1)

# <u>UNIT I</u>

Introduction: Biological data types, collection, classification schema of biological databases; Biological databases retrieval systems; Sequence and molecular file formats.

# <u>UNIT II</u>

Biological databases: Nucleotide database, protein database, structural database, genome databases, metabolic pathway database, literature database, chemical database, gene expression database, crop database with special reference to BTISNET databases.

# <u>UNIT III</u>

Bioinformatics Tools: Concept of alignment, scoring matrices, alignment algorithms, heuristic methods, multiple sequence alignment, phylogenetic analysis, molecular visualization tools.

# Practical

NCBI; Expasy: SwissProt; EBI; Search engines: ENTREZ and SRS; Perform local alignment using all BLAST variants; Multilple sequence alignment using ClustalW; T Coffee; phylogenetic analysis by PHYLIP; MEGA.

# **Suggested Readings**

Baxevanis AD, Ouellette BFF. 2001. Bioinformatics: A practical guide to the analysis of genes and proteins. John Wiley and Sons.

Mount DW. 2001. Bioinformatics: Sequence and Genome Analysis. Cold Spring Harbor. Xiong J. 2006. Essential Bioinformatics. Cambridge University Press.

<b>Course Title</b>	:	Structural Bioinformatics
Course No.	:	BTI/BTR 329
<b>Credit Hours</b>	:	3 (2-0-1)

# <u>UNIT I</u>

Introduction to structural databases of macromolecules, natural and synthetic small molecules; Structure of amino acids; Protein structure classification, Ramachandran plot; Experimental structure determination methods; Motifs, domain, profiles, fingerprint and protein family databases.

## <u>UNIT II</u>

Structural features of RNA, RNA secondary structure predictions; RNA folding; Small RNA prediction.

## <u>UNIT III</u>

Structure prediction: Basics of protein folding, protein folding problem, molecular chaperons; Secondary structure prediction methods and algorithms: Homology, *ab initio* and folding based tertiary structure prediction; Structure validation tools, energy minimization techniques; Introduction to molecular dynamics and simulation, Monte-Carlo methods, Markov chain and HMM; Structure visualization and comparison methods.

# Practical

Protein structural classification databases, 3D-Structural databases searching and retrieval, Ramchandran Plot, Structural visualization tools, Tools for protein secondary and tertiary structure prediction; RASMOL, Cn3D, CHIMERA, SWISSPDBviewer, CPH, MODELLER, SWISS Model, EasyModeler, Procheck; GROMAC; SANJIVNI; BHAGIRATH.

# **Suggested Readings**

A.Malcolm Campbell & Laurie J.Heyer. 2007. *Discovering Genomics, Proteomics and Bioinformatics*. Benjamin Cummings.

Allan Hinchkliffe. 2008. Modeling for Beginners. Wiley.

Creighton TE. 1993. Proteins: Structures and Molecular Properties. W.H. Freeman

Mount DW. 2001. Bioinformatics: Sequence and Genome Analysis. Cold Spring Harbor.

Setubal Joao & Meidanis Joao. 1997. Introduction to Computational Molecular Biology. PWS Publishing Company

<b>Course Title</b>	:	Pharmacogenomics
Course No.	:	<b>BTI/VPT 330</b>
<b>Credit Hours</b>	:	3 (2-0-1)

### <u>UNIT I</u>

Basic concepts of pharmacogenomics, clinical application and challenges in pharmacogenomics; Human Genome Project, genetic diseases, personalized medicine and pharmacogenomics necessity in drug designing; Prediction of structural changes among sequence variants and genetic analysis; Microsatellites for studying genetic variations; Drug databanks; Gene therapy.

## UNIT II

Drug Design: Study of important drug targets and their variations; Pharmacophore designing, prediction of ADME properties; Computational tool for toxicity prediction; SAR and QSAR techniques in drug designing; Drug receptor interactions; Structural based drug design; Lipinski's rule in drug design.

# Practical

Receptor-Ligand interactions, Pharmacophore development; OSDD; DrugBank; PubChem; molecular representation using SMILES; Chemsketch: 2D and 3D structure; Structure analyses using Chimera/VMD; Detection of active site of proteins using various software; bioavailability using Mol inspiration; Docking using HEX and AUTODOCK.

# **Suggested Readings**

Allan Hinchkliffe. 2008. *Modeling for Beginners*. Wiley- Blackwell Publishing. GerdFolkers, Wolfgang Sippl, Didier Rognan & Hans Dieter. 2003. *Molecular Modeling* 

*Basic Principles and applications.* Science. Gupta S.P. 1996. *Quantum Biology.* New Age. Lisa B. *Combinatorial Library Methods and Protocols* 

<b>Course Title</b>	:	Metabolomics and System Biology
Course No.	:	<b>BTB/BAS 327</b>
<b>Credit Hours</b>	:	3 (2-0-1)

## <u>UNIT I</u>

Metabolomics overview, major metabolic pathways: Glycolysis, Kreb's cycle, oxidative phosphorylation, amino acid, fatty acid and nucleotide metabolism, their control and integration; Metabolic flux and metabolic profiling; Catalytic mechanisms and enzyme kinetics, Michaelis-Menton kinetics; Conformational change, allosteric regulations, regulation of metabolic pathways; Signal transduction: Inter and intra cellular communications; Receptor ligand interaction; Structural components of signal pathways: G-protein, Jak-stat, receptor tyrosine kinase.

#### UNIT II

Signal Flow: Pathway to networks, small scale system biology experiments; System analysis of complex diseases, system pharmacology; Assembling large data sets in genomics and proteomics, computational analysis of large data sets, building networks; Mathematical representation of cell biological system, time and space.

# Practical

Metabolic pathway databases KEGG, BRENDA, Biosilico, Protein-protein interaction databases, Swiss 2D PAGE, E-PCR; Creating networks using Cytoscape, DAVID, MAS3; in silico functional annotation using GO, AGRIGO, PANTHER, BLAST2GO.

#### Suggested Readings

Berg JM, Tymoczko JL & Stryer L. 2002. *Biochemistry*. 5<sup>th</sup> Ed. W.H. Freeman and Company.

Fersht A. 1999. Structure and Mechanism of protein science. W.H. Freeman and Company.

Klipp E, Herwig R, Kowald A, Wierling C, Lehrach H. 2006. Systems Biology in practice. Concepts, implementation and Application. Wiley VCH.

Vaidynathan S, Harrigan GG, Royston Goodacre. 2005. *Metabolome analysis: Strategies for system biology*. Springer.

Voet D & Voet J. 2002. *Biochemistry*3<sup>rd</sup> Ed. John Wiley and Sons.

#### **SEMESTER-VII**

<b>Course Title</b>	:	<b>Educational Tour</b>
Course No.	:	BTS 411
<b>Credit Hours</b>	:	1 (0-1-0)

Local, neighboring or other University/SAU/Research Station/KVK/ICAR/ Institutes/ Industries etc. visit based on their importance and relevance for improvement of student exposure towards different Academic/ Industrial environments. With respective Advisor individually or associated group.

<b>Course Title</b>	:	Student READY In-house Skill Development Modules
Course No.	:	BTS 412
<b>Credit Hours</b>	:	20 (0-0-20)

Four Modules (Only one to be opted as per chosen elective) to be done with Advisor (Chairman) and 1/2 faculty engaged in corresponding modules (0+20).

- 1. Plant Biotechnology
- 2. Animal Biotechnology
- 3. Microbial and Environmental Biotechnology
- 4. Bioinformatics

### **Evaluation Criteria:**

Activity within each Module	Marks Distribution (%)
Attendance	10
Mid-term	25
Practical Assignments	15
Final Exam for Practical Skill Assessment	40
Oral Examination	10

<b>Course Title</b>	: Bioj	process Engineering, Bioseperation and Downstream processing
Course No.	:	BTO/AGE 411
<b>Credit Hours</b>	:	3 (3-1-0) [01 Tutorial of three contact hours per week]

## UNIT I

**Bioprocess Engineering:** Overview of a bioprocess including upstream and downstream processing. Chemical engineering principles applied to biological system, Principle of reactor design, ideal and non-ideal multiphase bioreactors, Mass and Heat transfer; Rheology of fermentation fluids, Aeration and agitation; Media formulation and optimization; Kinetics of microbial growth, substrate utilization and product formation; Sterilization of air and media; Batch, fed-batch and continuous processes; Various types of microbial and enzyme reactors; Instrumentation control and optimization; UNIT II

Heat Transfer Operations: Thermodynamics: Introduction to thermodynamics Systems; Closed, adiabatic, isolated, open and homogenous system, intensive, extensive and specific properties, thermodynamics state and equilibrium. Temperature and zeroth law of thermodynamics. Energy conservation and Temperature. Pressure; Density, specific weight, specific volume and specific gravity, Energy; Internal energy, kinetic energy, potential energy, process, cycle. Different laws of thermodynamics. Modes of Heat Transfer (Conductive, Connective and Radiative), heat transfer with change of phase, Governing Equation and Boundary conditions of heat transfer. Mass Transfer Operations: Equillibrium (Gas and Liquid, Gas and Solid, Solid and Liquid), Mass Conservation and Kinetics (Rate laws of homogeneous reactions and effect of temperature); Modes of Mass Transfer (Porous media flow, diffusive, Dispersive and convective), steady and unsteady state, Comparison of the mass transfer, Governing equations and Boundary conditions of Mass Transfer. Fluid Mechanics: Introduction, fluid properties, fluid statistics, manometers, static forces on immersed and curved surfaces, buoyancy, fluid kinematics, conservation of mass, stream function and velocity potential flow net, Euler's, Bernoulli's, momentum and angular momentum equations, kinetic energy and momentum. Dimensional analysis- Rayleigh's method and Buckingham Pi theorem, dimensionless numbers, model scales. Pipe, flow energy losses, Darcy-Weibach equation, estimation of friction factor, minor loses, pipe flow computations, hydraulic gradient and total energy line. Open channel flow, velocity distribution, uniform flow, flow resistance, flow computations, flow measuring devices. UNIT III

**Downstream Processing:** Upstream production and downstream; Unit operations in solid-liquid separation (filtration, centrifugation, settling, sedimentation, decanting and micro filtration), Cell Rupture, Recovery (Extraction- liquid-liquid, Adsorption) and Purification (Sedimentation, Precipitation, chromatography, electrophoresis and Membrane separation- reverse osmosis, electro dialysis and ultra filtration); Emerging separation techniques: Dynamic immobilization, reverse osmosis, super critical fluid extraction evaporation, super liquid extraction and foam based separation. Separation of intracellular, extracellular, heat and photosensitive materials. Product recovery trains - a few examples. Bioprocess design and development from lab to industrial scale; Microbial, animal and plant cell culture platforms; Production of biomass and primary/secondary metabolites; Biofuels, Bioplastics, industrial enzymes, antibiotics;

Large scale production and purification of recombinant proteins; Industrial application of chromatographic and membrane based bioseparation methods; Immobilization of biocatalysts (enzymes and cells) for bioconversion processes; Bioremediation-Aerobic and anaerobic processes for stabilization of solid / liquid wastes. Anaerobic and aerobic treatment of effluents. Typical examples for downstream processing and effluent disposal in process industries.

#### **SEMESTER-VIII**

<b>Course Title</b>	:Stud	lent READY Project Formulation, Execution and Presentation
Course No.	:	BTS 421
<b>Credit Hours</b>	:	10 (0-0-10)

Student READY - Project Formulation, Execution and Presentation 0+10

Evaluation Criteria: (Offered by Respective Advisor)

Parameter	Marks Distribution (%)
Project Formulation and Implementation	20
Work Performance	30
Regularity, General Conduct and Discipline	10
Initiative and Creativity	10
Final Presentation	15
Final Project Report	15

# Course Title: Student READY Entrepreneurial Development in BiotechnologyCourse No.: BTS 422Credit Hours: 10 (0-0-10)

#### Student READY - Entrepreneurial Development in Biotechnology 0+10

**OBJECTIVES:** Engage students in an innovative "actual hands on" learning experience in National/International industrial development. Challenge students to apply knowledge and skills, learned in the classroom, to real world problems in form of new product development. Enhance students' understanding of development processes, applications and complexities in Country, Industrial and emerging Markets contexts. Develop case studies and other learning materials (presentations etc.) to enrich classroom learning.

Various topics may be provided viz. Micropropagation; DNA fingerprinting; Genetic purity for maintenance breeding; Marker assisted selection; Haploid production; Database Management skills; Molecular Diagnostics; Recombinant protein production; Animal cell culture and maintenance; Fermentation, Biopharma production; Bioprocess enrichment; Bioremediation; Biofules, topics from respective module etc.

Evaluation Criteria: (	Offered by Respective Advisor)	

Parameter	Marks Distribution (%)	
Project Planning, Writing and Presentation	20	
Regularity	10	
Monthly Assessment	10	
Output Delivery	15	
Technical Skill Development	15	
Entrepreneurial and Business Networking Skills	10	
Report Writing	10	
Final Presentation	10	